Ecological Facets of Varied Plant Life Forms and their Therapeutic Relevance

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ABSTRACT

Life forms depict the sum of adaptive features attributed to the ecological relationships of species. Adaptation plays a crucial role in the natural selection of an organism and sustains an organism's survival in its habitat. These are evolutionary changes inherited by plants for their survival in nature and to cope with competition and stresses in the environment. Varied plant life forms exist in nature and are classified by Eugen Warming considering their corresponding traits and the associated response to the environment viz. hydrophytes, helophytes, oxylophytes, psychrophytes, halophytes, lithophytes, psammophytes, chersophytes, eremophytes, psilophytes, sclerophyllous, coniferous and mesophytes. These adaptive life forms bring about structural and physiological changes and evolve for the successful survival of organisms in nature. Secondary metabolites, fungal endophytes and phytohormones play a vital role in the adaptation process by providing them a defensive shield against stressors. When explored therapeutically, these adapted plants containing secondary metabolites contributed to preventing and treating a wide array of diseases and disorders and seem promising sources of drug discovery.

Keywords: Adaptation, Life forms, Secondary metabolites, Warming.

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INTRODUCTION

Greek philosopher and polymath, Aristotle (384-322) classified plant life forms considering their habitat and general appearance. Later on, his pupil, Theophrastus (371-287 BC) laid the concept of plant life forms by mentioning the botanical description and uses of several species in his famous writings. Carl Linnaeus (1707-78) strengthened this concept by recognizing algal groups along with vascular life forms and proposed a biological classification system.^[1] Although the term 'life form' came into light from the Aristotle period but was originally coined by Eugenius Warming in 1896.^[2] Danish botanist Christen Raunkier's established and further strengthened the fundamentals of the applied life form scheme.^[3] Plants possess an inherent ability to interact with biotic and abiotic agents and can survive under the extremities of nature.^[4] Plant adaptation is an extraordinary characteristic of a plant that facilitates it to survive under peculiar conditions of corresponding habitat.^[5] Limiting factors viz. food, competition, temperature, altitude and sunlight have a great impact on the ecological profile of an organism. Around 1895, a Danish botanist Eugenius Warming, studied the importance of these limiting



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factors on vegetation in ecology and classified plants into various ecological groups on the criteria of water requirements and the nature of substrate on which a particular species flourish.^[6] Warming, in his book 'Oecology of Plants- An Introduction to the study of plant communities' mentioned about oecological classification of plants (Table 1, Figure 1) and categorizing plants into several classes from Class 1 (section IV) to Class 13 (section XVI) viz. class 1 (hydrophytes), class 2 (helophytes), class 3 (oxylophytes), class 4 (psychrophytes), class 5 (halophytes), class 6 (lithophytes), class 7 (psammophytes), class 8 (chersophytes), class 9 (eremophytes), class 10 (psilophytes), class 11 (sclerophyllous), class 12 (coniferous), class 13 (mesophytes).^[7] Later on, this categorization of plants was modified and refined by various scholars due to advancements in our understating of ecological traits. These life forms undergo structural and physiological changes for their survival. Several morphological traits of these adaptive plants are depicted in Figure 2A.

Hydrophytes

Plants that dwell in water and wet places partially or wholly submerged are known as hydrophytes. Concerning their relationship with air and water, they can be classified into amphibious, floating and submerged types. Several adaptations found in hydrophytes include thin leaves, long slender, flexible stems having lacunae (submerged plants) and waxy cuticles.^[6] Examples include *Eichhornia crassipes*,^[8] *Ceratophyllum* *demersum*,^[9] *Polygonum senegalensis*,^[10] *Nymphaea lotus*,^[11] *Typha domingensis*.^[12]

Helophytes

Generally regarded as Marsh plants, helophytes are among the category of aquatic plants, whose assimilatory organs are submerged in water or soil or at most swim on the surface of water. They have reduced leaf surface, adventitious roots and horizontal rhizomes attributed to their adaptability. Examples are *Typha latifolia, Phragmites australis.*^[13]

Oxylophytes

Plant formations on sour/acidic soil are generally termed Oxylophytes. They are acid-loving plants adapted to free humous acidic soil. Well-developed coating of hairs, papillae, wax and thick cuticles are among the most common adaptive features of these plants e.g. *Vaccinium uliginosum*.^[7]

Psychrophytes

These plants are adaptable in cold and wet or excessively wet habitats represented by varied life forms viz. shrubs, under shrubs, sub-shrubs and perennial grasses. These plants are usually xeromorphic having stiff leaves with hard epidermis. Examples include *Ranunculus glacialis*,^[14] *Saxifraga flagellaris*.^[15]

Halophytes

They possess unique adaptive features viz. regulation of osmotic pressure and homeostasis, excretion via salt glands, altered membrane compositions, etc. which help them to tolerate high saline conditions.^[16] A few examples of halophytes are *Aerva javanica*, *Kochia scoparia*,^[17] *Acacia modesta*,^[18] *Rhizophora mucronata*,^[19] *Salicornia brachiata*.^[20]

Lithophytes

Rock-dwelling plants are often termed 'lithophytes' which generally obtain their nutrition from decomposing plant litter from surroundings, dew, rainwater, etc. They possess haptera by which they attach themselves to rocky surfaces and can also tolerate high calcium levels.^[7,21] Some examples of these rock-dwelling species are *Selaginella bryopteris*,^[22] *Pteris vittata*,^[23] *Dendrobium nobile*.^[24]

Psammophytes

These sand dune plants possess reduced leaves and exposed root systems to combat the extremities of gravel and sand.^[25] Examples include Agriophyllum squarrosum,^[26] Alhagi sparsifolia, Artemisia ordosica,^[27] Artemisia scoparia,^[25] Canavalia rosea,^[28] Digitaria ciliaris,^[29] Setaria vindis,^[30] Vitex trifolia.^[31]

Chersophytes

These are the plants that dwell on wasteland and play a crucial role in preventing soil erosion, as well as soil stabilization e.g. *Poa bulbosa*.^[7]

Eremophytes

Plants dwell on the desert and steppe remarked as eremophytes. Reduction in leaf size, sunken stomata, strong cuticle and thick-walled epidermis are among a few adaptive features of these plants^[7] e.g. *Haloxylon ammodendron, Nitraria sibirica*.^[27,32]

Psilophytes

They resemble chlorophyta (filamentous green algae) having true stems, sporangia, slender and branching rhizomes.^[33]

Sclerophyllous

Vegetation adapted to long stress periods of heat and dryness encompassing hard leaves and short internodes.^[34] Examples include *Eucalyptus camaldulensis*.^[35]

Coniferous

Plants having needle-like evergreen leaves, strong cuticularization of the epidermis and cone-bearing seeds are among the most distinguishable adaptive features of coniferous.^[36] Some commonly found coniferous plants are *Pinus roxburghii*, *Abies alba*.^[37]

Mesophytes

Plants that flourish in moderate or average water requirements/ conditions are termed Mesophytes. The primary distinguishing characteristics of this species are its well-developed roots with root caps, straight and branched branches, thick bark and waxy cuticles.^[38] Solanum lycopersicum^[39] and Biophytum sensitivum^[40] are a among few examples of mesophytes.

MULTIPLE FACETS OF LIFE FORMS

It has been observed that plants that were present in one life form can thrive in another. The most prominent example in this regard is of orchids, which are found to be inhabited in different life forms viz. terrestrial, lithophytic, epiphytic and saprophytic.^[41] This adaptive feature is also observed in Araceae,^[42] Lentibulariaceae^[43] and Orchidaceae.^[44] Danish Botanist, JEB Warming (1841-1924) mentioned the formations on the rocks (lithophytes) in oecological classification Section VIII, Class 6. The term 'lithophyte' was coined by Schimper in around 1898 representing the vegetation found on rocks or stones, whereas the term 'chasmophyte' was used for plants dwelling on crevices in rocks.^[45] Lithophytes adapted to rocky substrates and generally obtained their nutrition from the atmosphere.^[46] It is quite fascinating that many plants are adapted over time and able to flourish in distinct habitats representing multiplicity in their

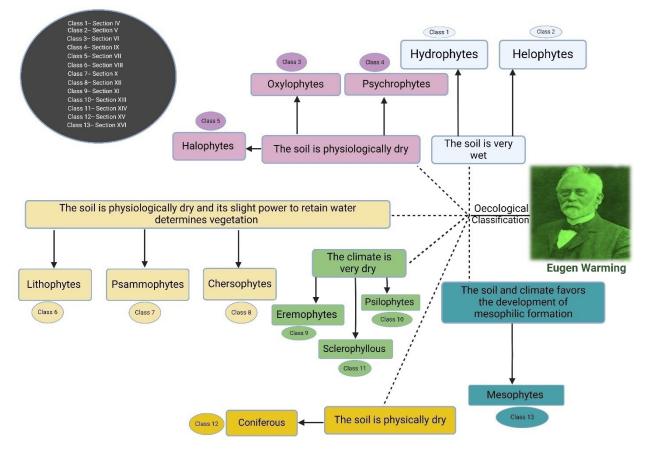


Figure 1: Overview of oecological/ecological classification.

Table 1: Warmings Oecological classification.^[7]

Class	Soil condition	Туре	Formations on	Section
1	The plant has access to plenty of water and the soil is	Hydrophytes	Water	IV
2	extremely wet.	Helophytes	Marsh	V
3	Physiologically dry soil and only partially supplied with water	Oxylophytes	Acidic soil	VI
4	for plant utilization. Therefore, the formations are primarily	Psychrophytes	Cold soil	IX
5	made up of xerophilous type.	Halophytes	Saline soil	VII
6		Lithophytes	Rocks	VIII
7		Psammophytes	Sand, gravel	Х
8		Chersophytes	Wasteland	XII
9	The climate is very dry and determines the character of the	Eremophytes	Desert and steppe	XI
10	vegetation and also controls soil properties; formations are	Psilophytes	Savannah	XIII
11	also xerophilic.	Sclerophyllous	Bush, forest	XIV
12	The soil is physically dry.	Coniferous	Forest	XV
13	The soil and climate favors mesophilic formations.	Mesophytes	Forest	XVI

life forms. *Begonia alicida* exists in both terrestrial and lithophytic life forms,^[47] *Persicaria capitata* is usually found in terrestrial habitats but is also found to dwell on dry rocks.^[48,49] *Peristylus monticola*, an orchid also inhabited terrestrial and lithophytic life forms.^[50] *Coelogyne corymbose*, an epiphytic orchid, weeping fern (*Lepisorus krameria*), and *Peperomia macrostachya* exist in both epiphytic and lithophytic life forms.^[51-53] *Selliguea griffithiana*

and *Ficus tinctoria* exist in terrestrial and epiphytic forms^[54,55] whereas *Rhododendron lochiae* exists in three life forms viz. terrestrial, lithophytic and epiphytic^[56] as documented in Table 2. Not only plants but several lichens were also found to flourish on rocks/stones as well as exist in epiphytic form. Hammered shield lichen (*Parmelia sulcata*) is frequently inhabited on the oak bark of various trees but is also found in the saxicolous form.^[57]

Species	Life form					
	Terrestrial	Lithophytic	Epiphytic	References		
Begonia alicida	+	+	-	[47]		
Begoniaceae						
Persicaria capitata	+	+	-	[48,49]		
Polygonaceae						
Peristylus monticola	+	+	-	[50]		
Orchidaceae						
Coelogyne corymbosa	-	+	+	[51]		
Orchidaceae						
Lepisorus krameria	-	+	+	[52]		
Aspleniaceae						
Peperomia macrostachya	+	+	-	[53]		
Piperaceae						
Selliguea griffithiana	+	-	+	[54]		
Polypodiaceae						
Ficus tinctoria	+	-	+	[55]		
Moraceae						
Rhododendron lochiae	+	+	+	[56]		
Ericaceae						

Table 2: Some examples of plants exist in varied life forms.

(+) represent existence in particular life form, (-) means non-existence in respective life form.

CHEMICAL RELEVANCE

THERAPEUTIC RELEVANCE

PSM (Plant Secondary Metabolites), fungal endophytes and phytohormones are involved in the defense mechanism of plants and can help them withstand the extremities of nature and other stressors which further aids in adaptation^[58] as represented in Figure 2B. It has been reported that a higher number of secondary metabolites viz. saponins, carotenoids, flavonoids, anthocyanins, etc. were produced in case plants flourish under alkaline soil conditions.^[59] Nitrogenous compounds viz. alkaloids have a prominent role in building plant defense mechanisms against herbivory and pathogens.^[60] Volatile organic compounds,^[61] flavonoids,^[62] anthocyanins,^[63] iridoids, tannins and polyphenols have a significant role in plant adaptation.^[64]

Fungal endophytes have also been reported to modulate plant growth are involved in adaptations and facilitate evolutionary transitions in plants.^[65] Brassinosteroids, a plant steroidal hormone have an important role in plant adaptation in response to abiotic stress^[66] along with jasmonic acid which plays a critical role in tissue wounding.^[67] Nevertheless, the role of ethylene, abscisic acid and salicylic acid has already been well-established in abiotic stress tolerance.^[68]

Plants evolved via adaptation and showed a wide array of pharmacological activities due to the presence of secondary metabolites as summarized in Table 3. These secondary metabolites have also been reported to be involved in providing defense to organisms against various types of stressors as well as helping them to adapt to peculiar conditions.[58] Water hyacinth Eichhornia crassipes showed antibacterial and antifungal activities when evaluated using several microbial strains.^[8] Several extracts of common hornwort (Ceratophyllum demersum) exhibited wound-healing properties.^[69] In a study involving a hydrophyte, Polygonum senegalensis hydroalcoholic extract was found potential inhibitor of a-glucosidase thus proving its curative role in treating diabetes.^[70] Hydro-ethanolic extract from white Egyptian lotus (Nymphaea lotus) scavenges free radicals in DPPH, lipid peroxidation and reducing power assays.^[71] Aerial parts of southern cattail (*Typha domingensis*) encompass complex secondary metabolites that successfully decrease levels of total cholesterol and triglycerides in Wistar rats.^[72] Helophytes viz. Typha latifolia and Phragmites australis showed anti-inflammatory properties.^[73,74] Crude extract and polyphenolic fractions of Vaccinium uliginosum, an oxylophyte showed antioxidant activity.^[75] Psychrophytes Ranunculus glacialis and Saxifraga flagellaris exhibited hemolytic and anticancer activities respectively.^[76,77] Halophytes Aerva javanica and Salicornia brachiata were reported as promising

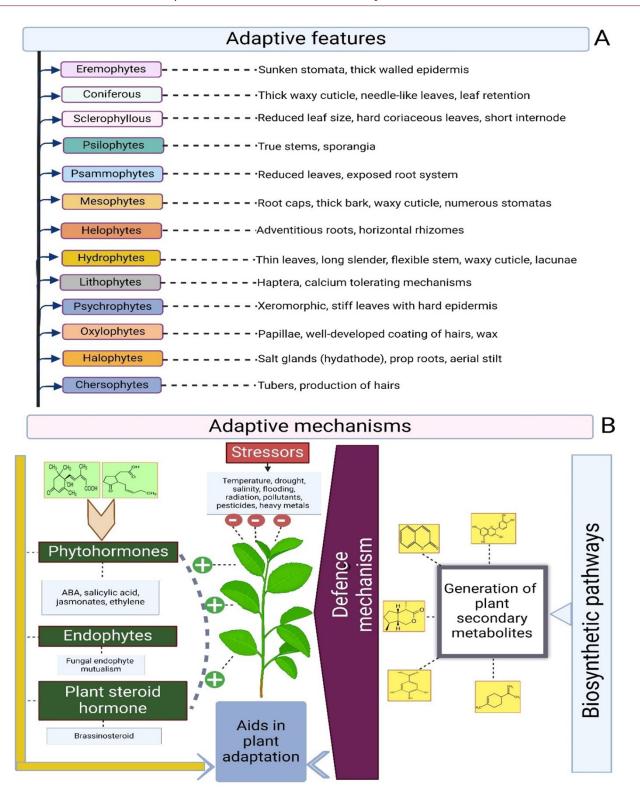


Figure 2: 2A) Adaptive features of plants, 2B) Adaptive mechanisms in plants.

antimicrobial plants.^[78,20] Mexican fireweed *Kochia scoparia* showed anticancer activities when evaluated in human breast cancer cells^[79] where as halophytes *Acacia modesta* and *Rhizophora mucronata* reported anti-inflammatory and antiviral activities respectively.^[80,19] *Selaginella bryopteris*, a lithophytic plant showed

its potential to treat stress.^[22] Fronds of lithophytic fern *Pteris* vittata showed wound-healing properties,^[23] whereas fragrant orchid *Dendrobium nobile* inhibited tumor.^[81] Psammophytes (*Agriophyllum squarrosum*, *Alhagi sparsifolia*) showed hepatoprotective potential.^[82,83] Artemisia ordosica, A. scoparia,

Table 3: Phytopharmacological status of adaptive life forms.
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SI. No	Plant	Part used	Phytoconstituents	Activity	References			
Hydro	Hydrophytes							
1	Eichhornia crassipes Pontederiaceae	Whole plant	Alkaloids, terpenes, phenolics.	Antibacterial, antifungal.	[8]			
2	Ceratophyllum demersum Ceratophyllaceae	Whole plant	Phenolics	Wound healing.	[69]			
3	Polygonum senegalensis Polygonaceae	Leaves	Saponins, flavonoids, tannins.	α-glucosidase inhibitory activity.	[70]			
4	Nymphaea lotus Nymphaeaceae	Leaves	Flavonoid	Antioxidant, anti-inflammatory.	[71]			
5	Typha domingensis Typhaceae	Aerial parts	Flavonoids, alkaloids, tannins, glycosides, polyphenols, coumarins, terpenes, saponins, proteins, carbohydrates.	Antihyperlipidemic.	[72]			
Helop	hytes							
6	Typha latifolia Typhaceae	Fruits	Polysaccharides	Wound healing.	[73]			
7	Phragmites australis Poaceae	Leaves	Polyphenols	Anti-inflammatory, antiviral.	[74]			
Oxylop	Oxylophyte							
8	Vaccinium uliginosum Ericaceae	Berries	Organic acids, anthocyanins, vitamins, glycosides.	Antioxidant.	[75]			
Psychr	ophytes							
9	Ranunculus glacialis Ranunculaceae	Stem, leaves	Volatile oils, phenolics	Hemolytic activity.	[76]			
10	Saxifraga flagellaris Saxifragaceae	Whole plant	Polyphenols	Anticancer, antioxidant, phytotoxic.	[77]			
Halop	hytes							
11	Aerva javanica Amaranthaceae	Leaves	Alkaloids, cardiac glycosides, steroids, terpenoids, resins.	Antibacterial.	[78]			
12	Salicornia brachiata Chenopodiaceae	Leaves	Alkaloids, flavonoids, tannins.	Antibacterial.	[20]			
13	Kochia scoparia Chenopodiaceae	Fruits	Triterpenoid glycosides, saponins.	Anti-cancer.	[79]			
14	Acacia modesta Fabaceae	Bark	Tannins	Anti-inflammatory, antipyretic, analgesic.	[80]			
15	Rhizophora mucronata Rhizophoraceae	Bark	Polysaccharides	Anti-viral.	[19]			
Lithop	Lithophytes							
16	Selaginella bryopteris Selaginaceae	Whole plant	Hexoses, proteins	Anti-stress, antioxidant.	[22]			
17	Pteris vittata Pteridoideae	Fronds	Polyphenols	Wound healing.	[23]			

SI. No	Plant	Part used	Phytoconstituents	Activity	References
18	Dendrobium nobile Orchidaceae	Entire herb	Alkaloids, bibenzyl, phenylpropanoids, phenanthrene, polysaccharides	Antitumor, anti-inflammatory.	[81]
Psamn	nophytes				
19	Agriophyllum squarrosum Amaranthaceae	Aerial parts	Oligosaccharides	Hepatoprotective.	[82]
20	Alhagi sparsifolia Fabaceae	Aerial parts	Flavonoids, alkaloids, polysaccharides, phenolic acids.	Antioxidant, hepatoprotective.	[83]
21	Artemisia ordosica Asteraceae	Root, stem, leaf	Dicaffeoylquinic acids.	Anti-inflammatory.	[84]
22	Artemisia scoparia Asteraceae	Aerial parts	Flavonoids,coumarins, chromones, phenolic acid, steroids,volatile oil.	Anti-inflammatory.	[85]
23	Digitaria ciliaris Poaceae	Flower	Xycaine, hexadecanoic acid, octadecanoic acid, linolenic acid, heptacosane, pentacosane.	Promote skin wound healing.	[86]
24	Vitex trifolia Lamiaceae	Leaf	Artemetin, casticin, vitexilactone, maslinic acid.	Anti-inflammatory.	[87]
25	Canavalia rosea Fabaceae	Leaves	Saponins, flavonoids, alkaloids, phlobatannins, cardiac glycosides.	Antimicrobial.	[88]
26	Setaria vindis Poaceae	Aerial parts	Tricin, p-hydroxycinnamic acid, vitexin 2"-O-glucoside.	Antioxidant.	[89]
Cherso	ophyte				
27	Poa bulbosa Poaceae	Stem, leaves	Alcohols, amines.	Antibacterial.	[90]
Eremo	ophytes				
28	Haloxylon ammodendron Amaranthaceae	Leaves	Quercetin, β-sitosterol, daucosterol	Antibacterial, antifungal.	[91]
29	Nitraria sibirica Nitrariaceae	Leaves	Flavonoids	Antioxidant, anti-proliferative.	[92]
30	Sophora alopecuroides Fabaceae	Aerial parts, seeds	Alkaloids, steroids, flavonoids, polysaccharides	Anticancer, antiviral, anti-inflammatory.	[93]
Sclero	phyllous				
31	Eucalyptus camaldulensis Myrtaceae	Leaves	Phenolics	Antimicrobial.	[94]
Conife	,				
32	Pinus roxburghii Pinaceae	Stem bark	Monoterpenes	Analgesic and anti-inflammatory.	[95]
33	Abies alba Pinaceae	Seed, cone	Essential oils	Antimicrobial, antiradical.	[96]
Meson	ohytes				

SI. No	Plant	Part used	Phytoconstituents	Activity	References
34	Biophytum sensitivum Oxalidaceae	Whole plant	Carbohydrates, proteins amino acids, flavonoids, saponins, tannins	Anti-urolithiasis.	[97]
35	Solanum lycopersicum Solanaceae	Fruit	β-carotene, vitamins, phenolics, carotenoids, glycoalkaloids	Anti-oxidant, anti-cancer.	[98]

Digitaria ciliaris, Vitex trifolia showed anti-inflammatory activities in various pharmacological models.^[84-87] *Canavalia rosea* and *Setaria vindis* possess antimicrobial and antioxidant properties.^[88,89] Silver nanoparticles synthesized from bulbous bluegrass (*Poa bulbosa*), a chersophyte showed antimicrobial potential.^[90] Organic extracts from several eremophytes viz. *Haloxylon ammodendron, Nitraria sibirica, Sophora alopecuroides* showed potent antimicrobial, antioxidant and anti-cancer activities respectively.^[91-93] Ethyl acetate extract from leaves of *Eucalyptus camaldulensis*, commonly known as river red gum showed antimicrobial and inhibitory action in schistosomiasis.^[94] Coniferous (*Pinus roxburghii, Abies alba*) and mesophytic plants (*Biophytum sensitivum, Solanum lycopersicum*) have already shown their well-established therapeutic potential when evaluated in different experimental models.^[95-98]

CONCLUSION

Adaptation seems to be an immensely invaluable gift that nature gave to organisms and proved necessary to bring evolutionary changes in them. The emergence of adaptive characteristics in an organism helped them to remain in harmony with the environment and for their optimum survival. Plant communities can adjust their life forms considering ecological factors and survive under the extremities of nature by undergoing modifications at structural and physiological levels. Several species also existed in distinct life forms by acquiring adaptive features in the evolutionary process and ultimately marked their importance from an ecological point of view viz. lithophytes and epiphytic. Secondary metabolites viz. polyphenols, flavonoids, iridoids and terpenoids emerged as protective agents in building the defense system of plants. Fungal endophytes and several phytohormones are also involved in the adaptation process and help adaptive life forms to withstand biotic and abiotic stresses. Adaptive life forms also encompassing a plethora of complex constituents emerged as a potential agent in the treatment of several diseases when tested experimentally using different pharmacological models. In this review, an attempt has been made to highlight multiple facets of the adaptive life form of plants and the role of agents involved in the process of adaptation.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

- 1. Fath BD. Encyclopedia of ecology. Netherlands: Elsevier Science; 2014.
- 2. Smith A. Bryophyte ecology. Springer Science+Business Media; 2012.
- 3. Raunkiaer C. The life forms of plants and statistical plant geography being the collected papers of C. Raunkiaer. United Kingdom: at the. Clarendon Press. 1934.
- Baucom RS, Heath KD, Chambers SM. Plant-environment interactions from the lens of plant stress, reproduction and mutualisms. Am J Bot. 2020;107(2):175-8. doi: 10.1 002/ajb2.1437, PMID 32060910.
- Shukla V, Kumar S, Kumar N. Plant adaptation strategies in changing environment. Singapore: Springer; 2017.
- 6. Shukla RS, Chandel PS. Plant ecology and soil science. S. Chand. 1994.
- 7. Warming E, Vahl M. Oecology of plants: an introduction to the study of plant-communities. Clarendon Press; 1909.
- Gebrehiwot H, Dekebo A, Annisa ME. Chemical composition, pharmacological activities and biofuel production of Eichhornia crassipes (water hyacinth): a review. J Turk Chem Soc Sect A Chem. 2022;9(3):849-66.
- Shaltout KH, Galal TM, El-Komi TM. Evaluation of the nutrient status of some hydrophytes in the water courses of Nile Delta, Egypt. Ecol Mediterr. 2010;36(1):77-87. doi: 10.3406/ecmed.2010.1377.
- 10. Metwally FE, Mohamed AA, Mahalel UA, Sheded MG. Evaluation of certain cosmopolitan hydrophytes in the Nile River, Aswan District for their ecological and bioactivity potentials: a review. Int J Sci Technol Res. 2020;9(1).
- Soliman MI, Ibrahim AA, Rizk RM, Naser NS. Phytoremediation, biochemical and molecular studies of some selected hydrophytes in Egypt. J Appl Sci. 2019;19(7):708-17. doi: 10.3923/jas.2019.708.717.
- Abbas AM, Lambert AM, Rubio-Casal AE, De Cires A, Figueroa EM, Castillo JM. Competition from native hydrophytes reduces establishment and growth of invasive dense-flowered cordgrass (*Spartina densiflora*). PeerJ. 2015;3:e1260. doi: 10.7717/pe erj.1260, PMID 26500809.
- 13. Krolikowska J. The transpiration of helophytes. Ekol Pol. 1978;26(2):193-212.
- Larcher W, Wagner J, Lütz C. The effect of heat on photosynthesis, dark respiration and cellular ultrastructure of the arctic-alpine psychrophyte *Ranunculus glacialis*. Photosynthetica. 1997;34(2):219-32. doi: 10.1023/A:1006840623763.
- 15. Chernov YI. The living tundra. United Kingdom: Cambridge University Press; 1988.
- Meng X, Zhou J, Sui N. Mechanisms of salt tolerance in halophytes: current understanding and recent advances. Open Life Sci. 2018;13(1):149-54. doi: 10.1515/ biol-2018-0020, PMID 33817080.
- Ferreira MJ, Pinto DC, Cunha Â, Silva H. Halophytes as medicinal plants against human infectious diseases. Appl Sci. 2022;12(15):7493. doi: 10.3390/app12157493.
- Khan M, Musharaf S, Shinwari ZK. Ethnobotanical importance of halophytes of Noshpho salt mine, District Karak, Pakistan. Res Pharm Biotechnol. 2011;3(4):46-52.
- Premanathan M, Kathiresan K, Nakashima H. Mangrove halophytes: A source of antiviral substances. South Pacific study. 1999;19(1-2):49-57.

- Manikandan T, Neelakandan T, Rani GU. Antibacterial activity of Salicornia brachiata, a halophyte. J Phytol. 2009;1(6).
- Green RT. Healthy and thrifty gardening helpful hints: A practical guidebook of 1001 wholesome living solutions to make life easier and save money with safe and natural nontoxic tips. CCB Publishing; 2012.
- 22. Sah NK, Singh SN, Sahdev S, Banerji S, Jha V, Khan Z, et al. Indian herb 'Sanjeevani' (Selaginella bryopteris) can promote growth and protect against heat shock and apoptotic activities of ultra violet and oxidative stress. J Biosci. 2005;30(4):499-505. doi: 10.1007/BF02703724, PMID 16184012.
- 23. Paul TS, Das BB, Talekar YP, Banerjee S. Exploration of the role of a lithophytic fern, *Pteris vittata* L. in wound tissue regeneration and remodelling of genes in hyperglycaemic rat model. Clin Phytosci. 2020;6:1-2.
- 24. Adhikari YP, Bhattarai P, Acharya KP, Kunwar RM, Bussmann RW. *Dendrobium nobile* Lindl. Orchidaceae. In: Ethnobotany of the Himalayas. Springer International Publishing; 2021.
- Huang WD, He YZ, Wang HH, Zhu YZ. Leaf physiological responses of three psammophytes to combined effects of warming and precipitation reduction in Horqin sandy land, northeast China. Front Plant Sci. 2021;12:785653. doi: 10.3389/fp ls.2021.785653, PMID 35058950.
- Chen G, Zhao J, Zhao X, Zhao P, Duan R, Nevo E, et al. A psammophyte Agriophyllum squarrosum (L.) Moq.: a potential food crop. Genet Resour Crop Evol. 2014;61(3):669-76. doi: 10.1007/s10722-014-0083-8.
- Xing-lei W, Piao-piao L, Min-jie Z, Jin-chao F, Ying L. Analysis of nutritional compositions and amino acid compositions of 6 common psammophytes in Alxa left Banner, Inner Mongolia. Spectrosc Spectral Anal. 2019;39(1):204-9.
- Araujo DD. Vegetation types of sandy coastal plains of tropical Brazil: a first approximation. In: Coastal plant communities of Latin America. Academic Press; 1992. p. 337-47.
- 29. Luo Y, Zhao X, Zhou R, Zuo X, Zhang J, Li Y. Physiological acclimation of two psammophytes to repeated soil drought and rewatering. Acta Physiol Plant. 2011;33(1):79-91. doi: 10.1007/s11738-010-0519-5.
- Qu H, Zhao H, Zhou R, Zuo X, Luo Y, Wang J, et al. Effects of sand burial on the survival and physiology of three psammophytes of Northern China. Afr J Biotechnol. 2012;11(20):4518-29.
- Shengfeng L, Lixiu R. Preliminary study of psammophytic vegetations on Poyang Lake side. J Plant Resour Environ. 1995;4(2):32-8.
- Jin ZZ, Zaynulla R, Lei JQ, Yakubov MA. Variation characteristics of water content in two typical eremophytes under drought stress in the drift desert hinterland. Appl Mech Mater. 2013; 316-317: 316-22. doi: 10.4028/www.scientific.net/AMM.316-317 .316.
- 33. Garber SD. Biology: a self-teaching guide. John Wiley & Sons; 2020.
- Bowman DM. Australian rainforests: islands of green in a land of fire. Cambridge University Press; 2000.
- Huang W, Tong YG, Yu GY, Yang WX. The sclerophyllous *Eucalyptus camaldulensis* and herbaceous Nicotiana tabacum have different mechanisms to maintain high rates of photosynthesis. Front Plant Sci. 2016;7:1769. doi: 10.3389/fpls.2016.01769, PMID 27933083.
- 36. VanCleave J. Janice VanCleave's super science models. John Wiley & Sons; 2004.
- Bhardwaj K, Silva AS, Atanassova M, Sharma R, Nepovimova E, Musilek K, et al. Conifers phytochemicals: A valuable forest with therapeutic potential. Molecules. 2021;26(10):3005. doi: 10.3390/molecules26103005, PMID 34070179.
- 38. Lack AJ, Evans DE. Plant biology. United Kingdom: Taylor & Francis; 2005.
- Zhou S, Palmer M, Zhou J, Bhatti S, Howe KJ, Fish T, et al. Differential root proteome expression in tomato genotypes with contrasting drought tolerance exposed to dehydration. J Am Soc Hortic Sci. 2013;138(2):131-41. doi: 10.21273/JASHS.138.2.1 31.
- Pawar AT, Vyawahare NS. Anti-urolithiatic activity of standardized extract of Biophytum sensitivum against zinc disc implantation induced urolithiasis in rats. J Adv Pharm Technol Res. 2015;6(4):176-82. doi: 10.4103/2231-4040.165017, PMID 26605159.
- Zhang S, Yang Y, Li J, Qin J, Zhang W, Huang W, et al. Physiological diversity of orchids. Plant Divers. 2018 Aug 1;40(4):196-208. doi: 10.1016/j.pld.2018.06.003, PMID 30740565.
- 42. Croat TB. Ecology and life forms of Araceae. Aroideana. 1998;11(3):4-55.
- Silva SR, Gibson R, Adamec L, Domínguez Y, Miranda VF. Molecular phylogeny of bladderworts: A wide approach of Utricularia (Lentibulariaceae) species relationships based on six plastidial and nuclear DNA sequences. Mol Phylogenet Evol. 2018;118:244-64. doi: 10.1016/j.ympev.2017.10.010, PMID 29054811.
- Weston PH, Perkins AJ, Entwisle TJ. More than symbioses: orchid ecology, with examples from the Sydney Region. Cunninghamia. 2005;9(1):1-5.
- 45. Moss CE. Vegetation of the Peak district. University Press; 1913.
- Zotz G. Plants on plants: the biology of vascular epiphytes. Germany: Springer International Publishing; 2016.
- Phutthai T, Sands M, Sridith K. Field surveys of natural populations of Begonia L. in Thailand. Thai Forest Bull (Botany). 2009;37:186-96.
- Pêgo RG, Oliveira LG, Garde GP, Grossi JA. Ornamental characteristics and vegetative propagation of *Persicaria capitata*. Acta Hortic International Symposium on New

Floricultural Crops 1000. 2013;VII(1000):(245-50). doi: 10.17660/ActaHortic.2013.10 00.32.

- 49. Dhaliwal DS, Sharma M. Flora of Kullu district. Bishen Singh Mahendra Pal Singh. 1999.
- Yeu NS, Nordin FA, Othman AS. Five new records of terrestrial and lithophytic orchids (Orchidaceae) from Penang Hill, Malaysia. Trop Life Sci Res. 2016;27(2):103-9. doi: 10. 21315/tlsr2016.27.2.8, PMID 27688854.
- Qin J, Zhang W, Zhang SB, Wang JH. Similar mycorrhizal fungal communities associated with epiphytic and lithophytic orchids of *Coelogyne corymbosa*. Plant Divers. 2020;42(5):362-9. doi: 10.1016/j.pld.2020.07.005, PMID 33134620.
- Negi S, Tewari LM, Pangtey YP, Kumar S, Martolia A, Jalal J, et al. Taxonomic studies on the family Polypodiaceae (Pteridophyta) of Nainital Uttarakhand. New York. Sci J. 2009;2(5):47-83.
- Fondom NY, Castro-Nava S, Huerta AJ. Seasonal variation in photosynthesis and diel carbon balance under natural conditions in two Peperomia species that differ with respect to leaf anatomy 1. J Torrey Bot Soc. 2009 Jan;136(1):57-69. doi: 10.3159/08-RA -085R.1.
- Lu HZ, Song L, Liu WY, Xu XL, Hu YH, Shi XM, et al. Survival and growth of epiphytic ferns depend on resource sharing. Front Plant Sci. 2016;7:416. doi: 10.3389/fpls.201 6.00416, PMID 27066052.
- Li Y, Mo YX, Cui HL, Zhang YJ, Dossa GG, Tan ZH, et al. Intraspecific plasticity and co-variation of leaf traits facilitate Ficus tinctoria to acclimate hemiepiphytic and terrestrial habitats. Tree Physiol. 2024; 44(2):tpae007. doi: 10.1093/treephys/tpae00 7, PMID 38198737.
- Craven LA, Withers RM. A second species of Rhododendron (Ericaceae) from Australia. Edinb J Bot. 1996;53(1):27-37. doi: 10.1017/S0960428600002699.
- Vondrák J, Liška J. Changes in distribution and substrate preferences of selected threatened lichens in the Czech Republic. Biologia. 2010;65(4):595-602. doi: 10.247 8/s11756-010-0061-3.
- Ramakrishna A, Ravishankar GA. Influence of abiotic stress signals on secondary metabolites in plants. Plant Signal Behav. 2011;6(11):1720-31. doi: 10.4161/psb.6.1 1.17613, PMID 22041989.
- Kumar M, Bharadwaj H, Kumari K. Plant adaptation to salinity stress: significance of major metabolites; 2023.
- Matsuura HN, Fett-Neto AG. Plant alkaloids: main features, toxicity and mechanisms of action. Plant Toxins. 2015;2(7):1-5.
- Boncan DA, Tsang SS, Li C, Lee IH, Lam HM, Chan TF, et al. Terpenes and terpenoids in plants: interactions with environment and insects. Int J Mol Sci. 2020;21(19):7382. doi: 10.3390/ijms21197382, PMID 33036280.
- Mouradov A, Spangenberg G. Flavonoids: a metabolic network mediating plants adaptation to their real estate. Front Plant Sci. 2014;5:620. doi: 10.3389/fpls.2014.0 0620, PMID 25426130.
- 63. Li Z, Ahammed GJ. Plant stress response and adaptation via anthocyanins: a review. Plant Stress. 2023;10:100230. doi: 10.1016/j.stress.2023.100230.
- 64. War AR, Taggar GK, Hussain B, Taggar MS, Nair RM, Sharma HC. Plant defense against herbivory and insect adaptations. AoB Plants. 2018; 10(4):ply037.
- Naranjo-Ortiz MA, Gabaldón T. Fungal evolution: major ecological adaptations and evolutionary transitions. Biol Rev Camb Philos Soc. 2019;94(4):1443-76. doi: 10.1111 /brv.12510, PMID 31021528.
- Ahammed GJ, Xia XJ, Li X, Shi K, Yu JQ, Zhou YH. Role of brassinosteroid in plant adaptation to abiotic stresses and its interplay with other hormones. Curr Protein Pept Sci. 2015;16(5):462-73. doi: 10.2174/1389203716666150330141427, PMID 25824388.
- Larrieu A, Vernoux T. QandA: how does jasmonate signaling enable plants to adapt and survive? BMC Biol. 2016;14:1-8.
- Larkindale J, Huang B. Effects of abscisic acid, salicylic acid, ethylene and hydrogen peroxide in thermotolerance and recovery for creeping bentgrass. Plant Growth Regul. 2005;47(1):17-28. doi: 10.1007/s10725-005-1536-z.
- Awati SS, Gilhotra RM, Singh SK, Raj V, Wadkar KA. Plant profile, phytochemistry and pharmacological properties of *Ceratophyllum demersum* Linn.: a review. SGVU J Pharm Res Educ. 2020;5(1):1-5.
- Bothon FT, Debiton E, Avlessi F, Forestier C, Teulade JC, Sohounhloue DK. *In vitro* biological effects of two anti-diabetic medicinal plants used in Benin as folk medicine. BMC Complement Altern Med. 2013;13:51. doi: 10.1186/1472-6882-13-51 , PMID 23452899.
- N'guessan BB, Asiamah AD, Arthur NK, Frimpong-Manso S, Amoateng P, Amponsah SK, et al. Ethanolic extract of Nymphaea lotus L. (Nymphaeaceae) leaves exhibits in vitro antioxidant, in vivo anti-inflammatory and cytotoxic activities on Jurkat and MCF-7 cancer cell lines. BMC Complement Med Ther. 2021;21(1):22. doi: 10.1186/ s12906-020-03195-w, PMID 33413340.
- 72. Akram A, Jabeen Q. Pharmacological evaluation of *Typha domingensis* for its potentials against diet-induced hyperlipidemia and associated complications. Trop J Pharm Res. 2022;21(3):563-9. doi: 10.4314/tjpr.v21i3.16.
- Gescher K, Deters AM. *Typha latifolia* L. fruit polysaccharides induce the differentiation and stimulate the proliferation of human keratinocytes *in vitro*. J Ethnopharmacol. 2011;137(1):352-8. doi: 10.1016/j.jep.2011.05.042, PMID 21669276.

- 74. Zhu L, Zhang D, Yuan C, Ding X, Shang Y, Jiang Y, *et al*. Anti-inflammatory and antiviral effects of water-soluble crude extract from *Phragmites australis in vitro*. Pak J Pharm Sci. 2017;30(4):1357-62. PMID 29039338.
- Kim YH, Bang CY, Won EK, Kim JP, Choung SY. Antioxidant activities of Vaccinium uliginosum L. extract and its active components. J Med Food. 2009;12(4):885-92. doi: 10.1089/jmf.2008.1127, PMID 19735191.
- Adeel H, Ahmad M, Rasool N, Mansha A, Bokhari T, Ullah A, et al. Biochemical and cytotoxic studies of various parts of *Ranunculus glacialis*. Oxid Commun. 2015;38(2).
- Ahmad I, Alam M, Wahab G, Habib I, Afzal S, Wasila H. Evaluation of anticancer, antioxidant and phytotoxic activities of (*Saxifraga flagellaris* Willd.). Pak J Agric Sci. 2022;59(5).
- Srinivas PS, Reddy SR. Screening for antibacterial principle and activity of *Aerva javanica* (Burm.f) Juss. ex Schult. Asian Pac J Trop Biomed. 2012; 2(2):S838-45. doi: 10.1016/S2221-1691(12)60321-9.
- Han HY, Kim H, Son YH, Lee G, Jeong SH, Ryu MH. Anti-cancer effects of *Kochia* scoparia fruit in human breast cancer cells. Pharmacogn Mag. 2014;10; Suppl 3:S661-7. doi: 10.4103/0973-1296.139812, PMID 25298688.
- Latif S, Ismail H, Khan MR, Rahim AA, Mehboob R, Dilshad E, et al. Pharmacological evaluation of Acacia modesta bark for antipyretic, anti-inflammatory, analgesic, antidepressant and anticoagulant activities in Sprague Dawley rats. Pak J Pharm Sci. 2020;33(3):1015-23. PMID 33191225.
- Zhang J, Xu HX, Zhao ZL, Xian YF, Lin ZX. Dendrobium nobile Lindl: a review on its chemical constituents and pharmacological effects. Chin Med Cult. 2021;4(4):235-42. doi: 10.4103/CMAC.CMAC 44 21.
- Bao S, Wu YL, Wang X, Han S, Cho S, Ao W, et al. *Agriophyllum* oligosaccharides ameliorate hepatic injury in type 2 diabetic db/db mice targeting INS-R/IRS-2/PI3K/ AKT/PPAR-y/Glut4 signal pathway. J Ethnopharmacol. 2020;257:112863. doi: 10.1016 /j.jep.2020.112863, PMID 32302715.
- Wei F, Yang X, Pang K, Tang H. Traditional uses, chemistry, pharmacology, toxicology and quality control of *Alhagi sparsifolia* Shap.: a review. Front Pharmacol. 2021;12:761811. doi: 10.3389/fphar.2021.761811, PMID 34721046.
- Xiao B, Wang JH, Zhou CY, Chen JM, Zhang N, Zhao N, et al. Ethno-medicinal study of Artemisia ordosica Krasch. (traditional Chinese/Mongolian medicine) extracts for the treatment of allergic rhinitis and nasosinusitis. J Ethnopharmacol. 2020;248:112262. doi: 10.1016/j.jep.2019.112262, PMID 31585162.
- Ding J, Wang L, He C, Zhao J, Si L, Huang H. Artemisia scoparia: traditional uses, active constituents and pharmacological effects. J Ethnopharmacol. 2021;273:113960. doi: 10.1016/j.jep.2021.113960, PMID 33636317.
- Park SM, Won KJ, Hwang DI, Kim DY, Kim HB, Li Y, et al. Potential beneficial effects of Digitaria ciliaris flower absolute on the wound healing-linked activities of fibroblasts

and keratinocytes. Planta Med. 2020;86(5):348-55. doi: 10.1055/a-1101-9326, PMID 32045946.

- Wee HN, Neo SY, Singh D, Yew HC, Qiu ZY, Tsai XC, et al. Effects of Vitex trifolia L. leaf extracts and phytoconstituents on cytokine production in human U937 macrophages. BMC Complement Med Ther. 2020;20(1):91. doi: 10.1186/s12906-020-02884-w, PMID 32188443.
- Vasanthi R, Balamurugan V. A review on pharmacological aspects of *Canavalia rosea*. Sci Progr Res. 2022;2(2):567-79. doi: 10.52152/spr/2022.172.
- Kwon YS, Kim EY, Kim WJ, Kim WK, Kim CM. Antioxidant constituents from Setaria viridis. Arch Pharm Res. 2002;25(3):300-5. doi: 10.1007/BF02976630, PMID 12135101.
- Allafchian A, Jalali SA, Mirahmadi Zare SZ, Amiri R. Biosynthesis of silver nanoparticles using *Poa bulbosa* extract and their antibacterial activity. Micro Nano Lett. 2022;17(13):349-55. doi: 10.1049/mna2.12139.
- Al-Hakimi AN, Alresheedi TM, Albarrak RA. The effect of the Saudi Haloxylon ammodendron shrub on silver nanoparticles: optimal biosynthesis, characterization, removability of mercury ions, antimicrobial and anticancer activities. Inorganics. 2023;11(6):246. doi: 10.3390/inorganics11060246.
- Chang Y, Lv G. Nitraria sibirica adapts to long-term soil water deficit by reducing photosynthesis, stimulating antioxidant systems and accumulating osmoregulators. Plant Physiol Biochem. 2024;206:108265. doi: 10.1016/j.plaphy.2023.108265, PMID 38091936.
- Wang R, Deng X, Gao Q, Wu X, Han L, Gao X, et al. Sophora alopecuroides L: an ethnopharmacological, phytochemical and pharmacological review. J Ethnopharmacol. 2020;248:112172. doi: 10.1016/j.jep.2019.112172, PMID 31442619.
- 94. Ghareeb MA, Habib MR, Mossalem HS, Abdel-Aziz MS. Phytochemical analysis of *Eucalyptus camaldulensis* leaves extracts and testing its antimicrobial and schistosomicidal activities. Bull Natl Res Cent. 2018;42:1-9.
- Kaushik D, Kumar A, Kaushik P, Rana AC. Analgesic and anti-inflammatory activity of *Pinus roxburghii* Sarg. advances in pharmacological and pharmaceutical sciences. 2012;2012(1):245431.
- 96. Wajs-Bonikowska A, Sienkiewicz M, Stobiecka A, Maciąg A, Szoka Ł, Karna E. Chemical composition and biological activity of *Abies alba* and *A. koreana* seed and cone essential oils and characterization of their seed hydrolates. Chem Biodivers. 2015;12(3):407-18. doi: 10.1002/cbdv.201400167, PMID 25766914.
- Pawar AT, Vyawahare NS. Anti-urolithiatic activity of standardized extract of Biophytum sensitivum against zinc disc implantation induced urolithiasis in rats. J Adv Pharm Technol Res. 2015;6(4):176-82. doi: 10.4103/2231-4040.165017, PMID 26605159.
- Chaudhary P, Sharma A, Singh B, Nagpal AK. Bioactivities of phytochemicals present in tomato. J Food Sci Technol. 2018;55(8):2833-49. doi: 10.1007/s13197-018-3221-z , PMID 30065393.

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