

Herbal Drug Development for Alzheimer's Disease: A Promising Approach through Computer-Aided Drug Design

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ABSTRACT

Dementia has arisen as a severe worldwide health concern in recent decades as the world's population ages. The most frequent kind of dementia is Alzheimer's Disease (AD). Despite substantial medication and developmental research, there are just a few FDA-approved medications for Alzheimer's disease. Alternative methods to AD therapies are urgently needed since existing medicines merely give symptomatic alleviation and are usually linked with side effects. A number of medicinal plants have been identified as nootropics, which increase mental and cognitive capabilities by influencing various brain physiological pathways. *In silico* methodologies, often known as Computer Aided Drug Design (CADD) investigations, are becoming more popular in industry and universities. They include a thorough grasp of molecular interaction from both a qualitative and quantitative standpoint. These approaches create and modify 3D molecular structures, distinct molecular attributes, and compute descriptors then design models, and use other computing drug research tools. The molecular structure of a system may be analyzed to extract useful information and forecast the potential of bioactive chemicals. This review focuses on *in silico* screening of certain phytochemicals which can be found beneficial in Alzheimer's disease.

Keywords: Alzheimer's disease, Alkaloids, Flavonoids, Terpene.

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INTRODUCTION

Alzheimer's Disease (AD) is an unalterable neurological illness that causes memory, behavior, visual-spatial perception, language, speech, cognition, and everyday functional activities to deteriorate over time, ultimately leading to death and dementia. Extracellular senile intracellular Neurofibrillary Tangles (NFT) and Neuritic Plaques (NP) are hallmarks of Alzheimer's Disease (AD)^[1] and may be seen in the brain parenchyma and brain arteries.^[2] Synaptic and neuronal function is disrupted, dendritic arborization are removed, and neurotransmitter levels are reduced, all of which contribute to gradual loss of neurons and brain capacity.^[3] Figure 1 contrasts the characteristics of a healthy brain to those of an AD patient in the advanced stages of the disease.

Neuritic plaques are beta-amyloid (A) protein aggregations (10–160m in diameter) surrounded by dead neurons, apolipoprotein E, astrocytes, microglia, proteoglycans, and α -1-antichymotrypsin.^[4] Three endoproteases, α -, β - and γ -secretase,^[5] sequentially degrade Amyloid Precursor Protein (APP) to create

A β peptides of various lengths.^[6] The primary isoforms of A β in plaques have become recognized to be A β 1-42, A β 4-42, and A β 1-40.^[7] According to the amyloid cascade theory, AD is caused by mutation in PSI, PSII, and APP genes that cause preferential APP cleavage, resulting in the formation of a longer variant of A peptide. These peptides have a propensity for forming sheets, which then clump together to create neuritic senile plaques surrounding neurons in brain tissue, resulting in synaptic damage and dendritic loss. Neuronal degradation and death occur as Alzheimer's disease advances, resulting in dementia and other signs.^[8]

The major component of intra-neuronal NFTs is tau protein, which is arranged in hyperphosphorylated paired helical strands.^[9] This has been hyperphosphorylated self-assembles into intermediate aggregates, coupled helical filaments, and NFTs.^[10] Due to the destabilization of the neuronal cytoskeleton, hyper-phosphorylation of this protein interrupts its normal activity in regulating vesicle-axonal transport, resulting in the loss of dendritic spines and the accumulation of neurofibrillary tangles, as well as toxic species of soluble tau, causing oxidative cytotoxicity, neurodegeneration, and inflammation.^[11]

In Alzheimer's disease, reactive oxygen species cause oxidative stress. Neurodegeneration is caused by oxidative stress, which causes free radical attacks on brain cells. However, O₂ is



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necessary for life, an unbalanced metabolism and excessive production of Reactive Oxygen Species (ROS) lead to various illnesses, like Parkinson's disease, AD, ageing, and a variety of other neurological illnesses. The histological and experimental data supporting the role of oxidants in the etiology of AD (oxidative-stress-hypothesis) is growing all the time.^[12]

One of most noticeable alterations in brains of Alzheimer's disease patients is a drop in hippocampal and cortical amount of the Acetylcholine (ACh) as disease progresses. On the behavioral side, a significant reduction in ACh levels impair cognitive performance.^[13] In several cholinergic routes in the peripheral and central nervous systems, AChE is engaged in the cessation of impulse transmission via fast hydrolysis of the neurotransmitter acetylcholine. Various inhibitors cause enzyme inactivation, which results in acetylcholine buildup, hyperstimulation of muscarinic receptors and nicotinic, and disruption of neuro-transmission. As a result, AChE inhibitors^[14] are used as relevant medications and poisons because they interact with the enzyme as their principal target.

This disease is progressive, which means that symptoms arise gradually and increase as time passes.^[15] Stages of Alzheimer's disease development have been defined based on suspected brain alterations and associated clinical changes.^[16] The alterations in the brain that cause Alzheimer's disease may start 20 years or more before symptoms manifest.^[17] Scientists refer to the period of time between the onset of AD and the onset of symptoms in advanced AD as the "continuum" of the disease. Despite these brain alterations, the person may operate properly at the start of the continuum. The brain's ability to adjust for neuronal injury has reached its limit which has happened as the person progresses along the continuum, and cognitive function begins to deteriorate. Afterwards, the death and damage of neurons is severe enough that the person exhibits clear signs of cognitive impairment, such as memory loss or confusion about time and location. Basic body processes such as swallowing become affected later, leading to death.^[18,19]

Synaptic plasticity and memory function are both controlled by the N-methyl-D-aspartate (NMDA) receptor. Extra glutamate may be produced from injured cells in Alzheimer's disease, resulting in persistent calcium overexposure, which may accelerate cell destruction.^[20]

Mitochondrial failure, oxidative stress, and reduced metabolism may all be linked to neurodegenerative diseases, like normal brain ageing.^[21] According to system biology, effective therapy of complex illnesses like Alzheimer's and cancer necessitates the restoration of broken disease networks, which often necessitates the simultaneous manipulation of numerous proteins (targets)/pathways.^[22] As a result, addressing numerous pathways might result in effective and useful treatment options for AD.

The restricted number of FDA-approved medications that are frequently used has made treating Alzheimer's disease difficult. Furthermore, these medications treat AD symptomatically, offering temporary comfort rather than treating the illness by improving its pathophysiology, and have been linked to a variety of side effects. As a result, there is a pressing need to create new anti-drugs.^[23]

Plant-based therapies for treating health problems have lately sparked a lot of attention, despite the fact that they have been utilized by humans since last many centuries. This old traditional practice of employing medicinal plants has resulted in a large body of anecdotal data demonstrating the safety and usefulness of several species. As a consequence of secondary metabolism, medicinal plants provide an infinite variety of chemicals, resulting in more chemical diversity than other natural sources. For AD medication development, researchers have demonstrated a keen interest in exploring historically used medicinal herbs, their compounds, and even their combinations.^[24]

Computer Aided Drug Design (CADD) is based on the assumption that pharmaceutically active chemicals interact with macro-molecule targets, most often nucleic acids or proteins. The steric complementarity of contacting surfaces of molecules, hydrophobic interactions, electro-static force, and the creation of hydrogen bonds are all important elements in such interactions. These parameters are mostly evaluated during the study and prediction of two-molecule interactions.

Flavonoids

Several scientific investigations^[25,26] have shown that polyphenols have beneficial properties. Their most apparent and simple activity is to reduce cell damage induced by free radicals, which has also been detected in Alzheimer's disease.^[27,28] Rodacka *et al.*^[29] discovered that the polyphenols resveratrol and tiron had neuroprotective effects through inactivating Glyceraldehyde-3-phosphate Dehydrogenase (GAPDH) produced by superoxide anion radicals. The docking of both ligands with the enzyme demonstrated that the radical's resveratrol and tiron are more effective than the superoxide anion alone in inactivating glyceraldehyde-3-phosphate dehydrogenase, with tiron being the greater antioxidant.

Utilizing examples from earlier scientific investigations using water extracts, Lakey-Beitia *et al.*^[30] demonstrated that extracts from *Caesalpinia crista* leaves inhibited A β aggregation from monomers and dissolved preformed A β fibril.^[31] *Ginkgo biloba* extract decreased oligomer formation,^[32] *Centella asiatica* decreased synuclein aggregation,^[33] and *Paeonia suffruticosa* extracts decreased A fibril production while also destabilizing preexisting amyloid fibrils.^[33] He hypothesized that polyphenols may modify APP processing, inhibit A β aggregation and inhibit fibrils. Polyphenolic glycosides like rutin, apigenin, and

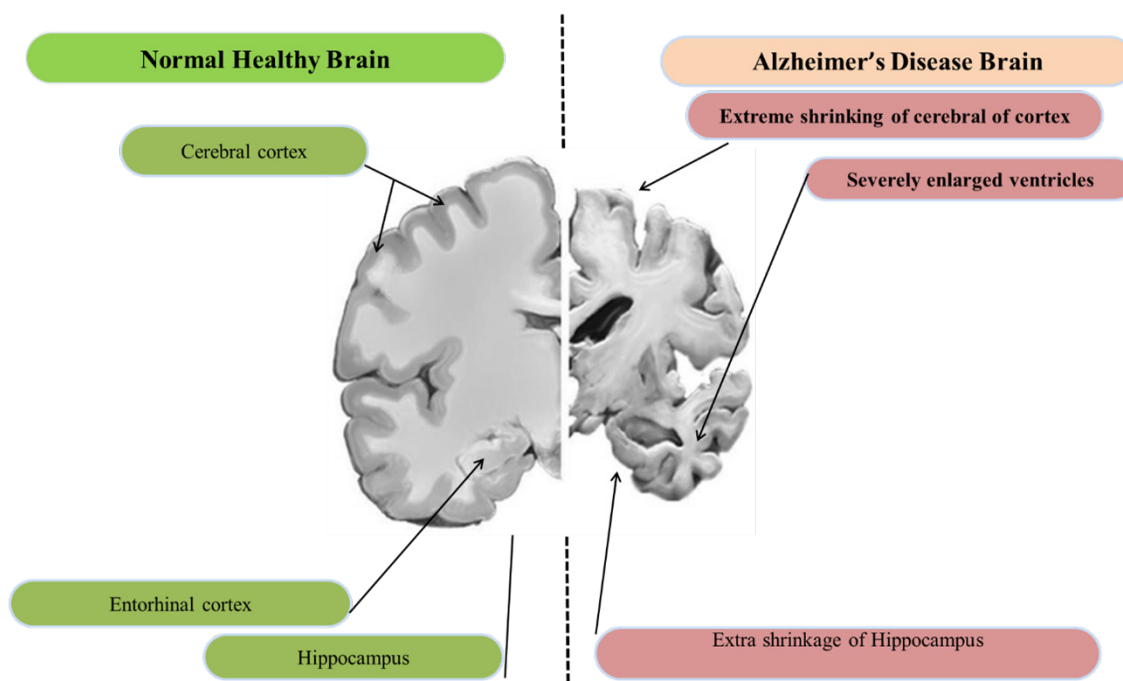


Figure 1: Anatomical changes in Normal brain vs Alzheimer's brain.

naringenin, according to Mora-Pale *et al.*,^[34] inhibit β -amyloid aggregation.

Wang *et al.* 2012^[35] investigated Mangostin, a polyphenolic xanthone produced from *Garcinia mangostana* Linn. This molecule has been used in various investigations^[36,37] and has been found to exhibit membrane-protective,^[38] anti-malarial,^[39] anti-viral,^[40] anti-bacterial,^[41] and other actions.

Tau proteins are the main components of glial fibrillar and intra-neuronal lesions in AD, which cause dementia and accelerate the neuro-degenerative process. Madeswaran *et al.*^[42] used molecular docking to investigate the tau protein kinase I inhibitory effect of six flavonoids. Catechin, acacatechin, scopoletin, galangin, memantine, and silbinin were the chemicals studied. Lipinski's rule was used to compute and assess parameters such as molecular weight, molecular formula, rotatable bonds, aromatic carbons, and the number of torsions. The lowest energies were found in galangin and silbinin, which might be employed in future research to treat Alzheimer's disease.

Six flavonoid compounds derived from the local medicinal plant *Rhus parviflora* (Anacardiaceae), quercetin-3-O-b-D-galactopyranoside, sulfuretin, ureusidin-6-O-b-D-glucopyranoside, aureusidin, cupressuflavone, and hovetrichoside C were shown to be CDK inhibitors in research by Shrestha *et al.*^[42] Molecular docking was used to test the chemicals. CDK5/p25 three-dimensional structures were retrieved from the RCSB protein data bank, while produced flavonoid compounds structures were chosen from the National Center for Biotechnology Information's Pubchem chemical database. Through hydrogen bonding with active-site residues at

GLN130 and CYS83, all of the flavonoids demonstrated excellent interactions with the receptor. Aureusidin and auroras, on the other hand, yielded higher energy values.

The top-class of medications used to cure AD were acetylcholinesterase inhibitors. Most often used AChE inhibitors, such as tacrine and physostigmine, have shown certain issues in clinical investigations, including difficulty with hepatotoxic potential and oral administration. The discovery of donepezil hydrochloride, which launched a new class of acetylcholinesterase inhibitors with longer and more selective action and tolerable side effects, was the result of research aimed at finding a new kind of acetylcholinesterase inhibitor. Sugimoto and coauthors used CADD investigations to find novel inhibitors, such as QSAR-3D, Comparative Molecular Field Analysis (CoMFA), and docking. Because there is a negative connection between IC_{50} and DE, QSAR modelling on the benzamine set shows that the cis isomers of the molecule have a higher intrinsic activity than the trans isomers (C-T).

As AChE inhibitors, researchers are now using secondary metabolites such as flavonoids, alkaloids, and xanthones. Yang and colleagues, for example, produced a series of aporphine alkaloid analogues. Ellman's technique (modified) was used to determine the anti-acetylcholinesterase activity of the produced compounds, as detailed in the work published in 2012. Molecular docking was used to find two highly active molecules (nuciferine derivatives) that interacted as ligands with the acetylcholinesterase site: 1, 2-dihydroxyaporphine and dehydronuciferine.^[43,44]

In AD, alterations in the cholinergic system are a key source of worry. The substrate is the most significant distinction between

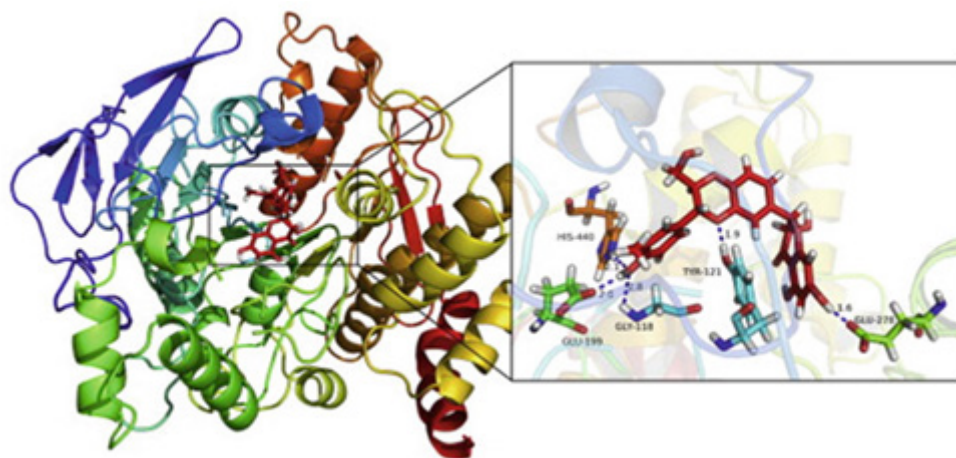


Figure 2: The combined conformation of the A β -silibinin complex shows that silibinin binds to the active sites of AChE.

the two cholinesterases: AChE hydrolyzes acetylcholine in the brain synapses and blood, while butyrylcholinesterase hydrolyzes Butyrylcholine (BChE) in the liver.^[45-47] Two forms of cholinesterase's breakdown acetylcholine, therefore inhibiting these both enzymes with a dual inhibitor could result in increasing the acetylcholine levels in the brain and improved therapeutic effect. Drug candidates such as rivastigmine, physostigmine, velnacrine, donepezil and tetrahydro aminoacridine (tacrine) are being studied as AChE inhibitors.^[48,49]

A most common pathological feature of AD is Amyloid plaques in the brain.^[50] A 39–43 amino acid peptide, the amyloid-peptide (A), is the plaques' most important protein component. A deposition produces Reactive Oxygen Species (ROS), that are implicated in the neurodegenerative and inflammatory pathophysiology of AD.^[51,52] The amyloid-peptide (A) produced by proteolytic cleavage of Amyloid Precursor Protein (APP) by β - and γ -secretases is a key component of amyloid plaques. Many writers ascribe cognitive decline to the deposition of amyloid plaque, which is a hallmark of Alzheimer's disease, although there is a suggestion that plaque production is a result of metabolic deterioration in AD patients' brains.^[53] According to recent research, Acetylcholinesterase (AChE) may increase formation of amyloid fibril by interacting with the PAS of acetylcholinesterase, which gives stability to the acetylcholinesterase-A complex. This is more hazardous than single A peptides alone.

Utilizing a spectrophotometric approach, Li *et al.*^[54] investigated various compounds against both cholinesterases, utilizing galanthamine and tacrine as reference compounds. Using curcumin as a reference, aggregation inhibition, self-induced kinetic ChE inhibition, and metal-chelating activities were investigated. An isolated compound showed to be the most powerful AChE inhibitor in these tests, thus the authors molecular docked it with TcAChE id PDB: 2ckm. Through π -contacts with distances of 4.24 and 5.05 Å, the flavonoid ring interacted between the indole ring of Tyr70 or Trp 279. Despite the fact

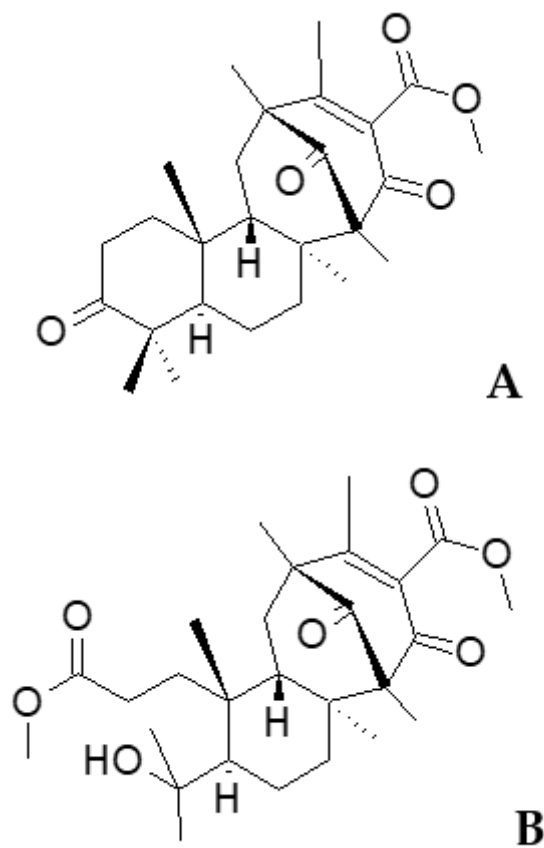


Figure 3: (A, B) Asperterpenes.

that 13 s is a powerful inhibitor, 13 k has more multipurpose capabilities.

Much research on multifunction anti-drugs Alzheimer's have been published, with some of them employing natural compounds.^[55-57] Lou *et al.*^[58] synthesized and tested 17 4-dimethylamine flavonoid derivatives for their anticholinesterase,

anti-A aggregation, and anti-ROS properties. The scientists used two enzymes from the Protein Data Bank to analyse the ligands, in this investigation π - π interactions were also discovered in the residue Trp 279. The findings revealed inhibitory potentials for both BChE and AChE that were comparable to or better than rivastigmine.

To clarify the structural characteristics of flavonoid by products which contribute to the inhibitory action of acetylcholinesterase, Goyal *et al.*^[59] used pharmacophore-based virtual screening and 3D Quantitative Structure-Activity Relationship (QSAR) to study 24 flavonoids. Using comprehensive docking experiments, the binding mechanisms of action with the enzyme were examined using a QSAR-3D model. Two aromatic rings, one hydrogen-bond acceptor, two hydrophobic region, and were the best pharmacophore hypotheses. 4 lead compounds were identified as a result of this research, one of which was shown to be particularly effective in inhibiting the dual AChE-binding site.

Chakraborty *et al.*^[60] used QSAR models to evaluate polyphenols' anti-amyloidogenic effects. CODESSA was given eighteen chemicals to calculate quantum chemical and thermodynamic descriptors for electrostatics, geometrical, quantum chemical, topological, constitutional, and thermodynamic properties. The best QSAR model was then used to create a database of 200 phytochemicals. This strategy is used to find fresh leads.

Morin is a flavonoid that has been shown to have anti-aggregation properties in A β monomers and dimers^[61] as well as inhibitory properties in the enzyme AChE.^[62] Morin's hydrogen-bonding capacity and aromaticity, hydrophobicity were detected in molecular dynamic simulations, and these properties may influence binding energies. Morin has an effect on A's tertiary and quaternary structure. Morin's AChE inhibition was explored using molecular docking, much as galangin, myricetin, fisetin, and quercetin. The structure activity correlations were then constructed by Remya and associates.^[63]

A flavonoid silibinin, derived from *Silybum marianum*, was investigated to see whether it may be utilized for the treatment of AD. It acts as a dual inhibitor of acetylcholinesterase and peptide aggregation. The researchers investigated the affinity of silibinin with A β and acetylcholinesterase *in silico* using molecular dynamics and docking simulations.^[64] The flavonoid connects with the active sites of acetylcholinesterase in Figure 2.

Alkaloids

Alkaloids are N-containing heterocyclics that are found in a variety of plant species. The majority of alkaloids are hazardous (including nicotine, strychnine, morphine, ephedrine, quinine), but researchers have found bioactive,^[65] including fungal infection treatment,^[66] anti-malarial,^[67] anti-HIV,^[68] and inhibition of AChE in Alzheimer's disease.^[69-72]

The isoquinoline, quinolizidine, triterpenoidal/steroidal, and indole classes of Amaryllidaceae, Lycopodiaceae and Buxaceae are regarded major alkaloid inhibitors of BChE and AChE, according to Konrath *et al.*^[73] They are described in terms of their structural variety and physico-chemical qualities, with a focus on SARs and docking investigations. The researchers demonstrate that a positively charged nitrogen interacts with one of the AChE-binding sites.

Another enzyme linked to Alzheimer's disease is Retinoblastoma-associated protein (RbAp48), which causes memory loss when deficient. Huang *et al.*^[74] used Perivine, eicosandioic acid, and bitter-sweet alkaloid II from traditional Chinese medicine in docking investigations.

Makowska *et al.*^[75] used molecular docking and hydrophilic interaction liquid chromatography to investigate trigonelline's affinity for the A(1-42) peptide. Trigonelline is associated with the His 6, 13 and 14, and Tyr10 residues in the A(1-42) protein, according to the researchers.

The alkaloids 3-epimacronine and lycoramine found in *Zephyranthes carinata* plant have been shown to inhibit BChE and AChE may be useful for the treatment of AD. Cortes *et al.*^[76] discovered that 3-epimacronine derivatives were selective AChE ligands in a docking investigation. Analytical experiments utilising GCMS and an assay evaluating inhibition of acetylcholinesterase activity revealed that alkaloids generated by other species in Amaryllidaceae family also inhibit BChE and AChE.^[77-80]

The Apocynaceae family's *Holarrhena anti-dysenterica* has a long history of usage in medicinal purposes, and its alkaloids have been shown to have anti-malarial, anti-bacterial, anti-gastro intestinal, and anti-cancer disease properties.^[81-84] Yang and colleagues^[85] looked examined conimin, conarrhimin, conessimin, isoconessimine, and conessine, which were all isolated from *H. anti-dysenterica*. The inhibitory action of AChE was investigated *in vitro*, and the authors used the data to create a SAR model. Docking with the enzyme was used to test the compound. The amount of methyl groups in a ligand improved its ability to inhibit AChE, and all ligands interacted with the protein's hydrophobic pocket.

Ul-Haq *et al.*^[86] investigated physostigmine analogues although the alkaloid physostigmine inhibits AChE,^[86-91] it has a short, varied bio-availability, half-life, and minimal clinical efficacy. The compounds were chosen from studies based on their AChE inhibiting activity IC₅₀ values (9.78–20893.1nM); the IC₅₀ values were transformed to pIC₅₀ values (logIC₅₀). The researchers employed 32 alkaloids for the training set and eight for the test set, and they were separated into two groups: alkyl and phenyl chains and ionizable nitrogen of the morpholine moiety. The energy minimization of compound structures was utilised to construct 3DQSAR (PLS regression analysis), CoMFA and CoMSIA models. The physostigmine analogues' steric and

electrostatic potentials, as well as their atomic co-ordinates, charges, physico-chemical properties (electrostatic, hydrophobic, steric, and hydrogen bond acceptor and donor) were evaluated using these tools. The models demonstrated significant predictive abilities as well as structure-activity correlations, facilitating and encouraging the use of these substances in the treatment of Alzheimer's disease.

Terpenoids

Dimethylallyl Pyrophosphate (DMAPP) and Isopentenyl Pyrophosphate (IPP) are the two major precursors for monoterpenes, polyterpenes, diterpenes, sesquiterpenes, tetraterpenes, and triterpenes. Which are subclassified by their structure: monoterpenes, sesquiterpenes. Despite the range of classes, these natural products are said to include a large number of chemicals and have a variety of beneficial effects,^[92-95] including anti-disease Alzheimer's;^[96-99] rational planning is not required.

A marine natural substance, furano-sesquiterpene palinurin, was isolated by Bidon-Chanal *et al.*^[100] Its inhibiting impact on glycogen synthase kinase 3b was discovered by the researchers (GSK-3b). The enzyme under investigation might be used to treat AD, neurological illnesses, myocardial ischemia, and cancer.^[101-104] The team identified two pockets in the receptor, one at the N-terminal lobe and second at the C-terminal lobe, as possible allosteric binding sites using molecular docking. The compound's inhibitory action is based on site binding.

Two asper terpenes derived from the culture broth of *Aspergillus terreus* were the first to be reported as potent inhibitors of BACE1. Amyloid fibril, KMO, GlpG, PSH, NMDA receptor, acetylcholinesterase, BACE1, and APOE4 were studied (Figure 3) using molecular docking with 8 important enzymes implicated in AD processes.^[105] BACE1 binds to ligands with a higher affinity over the various targets. Although having lower predicted binding energies.

CONCLUSION

Alzheimer's Disease (AD) is the most prevalent form of dementia, and it is now a worldwide and economic danger. Because Alzheimer's disease is a multifactorial disease, there are a variety of treatment targets. The FDA-approved medication for Alzheimer's disease now only addresses symptoms and does not address memory issues or slow the pace of AD neurodegeneration. As a result, there is still a pressing need to find alternative methods to AD treatments that target many underlying pathways in order to improve AD therapy. Many medicinal plants may be effective in the treatment of serious illnesses such as stroke, cancer, neurodegenerative disorders, and cardiovascular problems, according to traditional medical systems. Efficacy of several active substances obtained mostly from medicinal plants in dementia, particularly Alzheimer's disease, has been studied. In reality,

modern anti-medications, Alzheimer's like galantamine and rivastigmine, are phytochemical (alkaloids)-based medications that are selective, competitive AChE inhibitors that may be synthesised or derived from *Galanthus sp.* bulbs and flowers. Some Phyto drugs have recently been thoroughly investigated in cell culture and animal models of Alzheimer's disease, as well as in clinical studies. Extracts of *Curcuma longa*, *Ginkgo biloba*, *Angelica sinensis*, and *Withania somnifera* showed promise in the treatment of Alzheimer's disease and should be explored further to understand their maximum potential.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- de Leon BK, MJ, Zetterberg H. J Alzheimers Dis. 2006;387-403.
- Poulin P, Zakzanis KK. *In vivo* neuroanatomy of Alzheimer's disease: evidence from structural and functional brain imaging. Brain Cogn. 2002;49(2):220-5. PMID 15259395.
- Stadelmann C, Deckwerth TL, Srinivasan A, Bancher C, Brück W, Jellinger K, *et al.* Activation of caspase-3 in single neurons and autophagic granules of granulovacuolar degeneration in Alzheimer's disease: evidence for apoptotic cell death. Am J Pathol. 1999;155(5):1459-66. doi: 10.1016/S0002-9440(10)65460-0, PMID 10550301.
- Perry G, Zhu X, Smith MA, editors. Alzheimer's disease: advances for a new century. Los Press; 2013.
- Skovronsky DM, Fath S, Lee VM, Milla ME. Neuronal localization of the TNF α Converting Enzyme (TACE) in brain tissue and its correlation to amyloid plaques. J Neurobiol. 2001;49(1):40-6. doi: 10.1002/neu.1064, PMID 11536196.
- Chow VW, Mattson MP, Wong PC, Gleichmann M. An overview of APP processing enzymes and products. Neuro Molecular Med. 2010;12(1):1-12. doi: 10.1007/s12017-009-8104-z, PMID 20232515.
- Portelius E, Bogdanovic N, Gustavsson MK, Volkman I, Brinkmalm G, Zetterberg H, *et al.* Mass spectrometric characterization of brain amyloid beta isoform signatures in familial and sporadic Alzheimer's disease. Acta Neuropathol. 2010;120(2):185-93. doi: 10.1007/s00401-010-0690-1, PMID 20419305.
- Hardy J. The amyloid hypothesis for Alzheimer's disease: a critical reappraisal. J Neurochem. 2009;110(4):1129-34. doi: 10.1111/j.1471-4159.2009.06181.x, PMID 19457065.
- Imbimbo BP, Lombard J, Pomara N. Pathophysiology of Alzheimer's disease. Neuroimaging clinics. 2005;15(4):727-53.
- Spires-Jones TL, Stoothoff WH, de Calignon A, Jones PB, Hyman BT. Tau pathophysiology in neurodegeneration: a tangled issue. Trends Neurosci. 2009;32(3):150-9. doi: 10.1016/j.tins.2008.11.007, PMID 19162340.
- Gooch MD, Stennett DJ. Molecular basis of Alzheimer's disease. Am J Health Syst Pharm. 1996;53(13):1545-57; quiz 1603. doi: 10.1093/ajhp/53.13.1545, PMID 8809275.
- Behl C, Moosmann B. Antioxidant neuroprotection in Alzheimer's disease as preventive and therapeutic approach. Free Radic Biol Med. 2002;33(2):182-91. doi: 10.1016/s0891-5849(02)00883-3, PMID 12106814.
- Cummings JL. Treatment of Alzheimer's disease: current and future therapeutic approaches. Rev Neurol Dis. 2004;1(2):60-9. PMID 16400259.
- Colović MB, Krstić DZ, Lazarević-Pašti TD, Bondžić AM, Vasić VM. Acetylcholinesterase inhibitors: pharmacology and toxicology. Curr Neuropharmacol. 2013;11(3):315-35. doi: 10.2174/1570159X11311030006, PMID 24179466.
- Raber J. AR, apoE, and cognitive function. Horm Behav. 2008;53(5):706-15. doi: 10.1016/j.yhbeh.2008.02.012, PMID 18395206.
- Reddy PH, Tripathi R, Troung Q, Tirumala K, Reddy TP, Anekonda V, *et al.* Abnormal mitochondrial dynamics and synaptic degeneration as early events in Alzheimer's disease: implications to mitochondria-targeted antioxidant therapeutics. Biochim Biophys Acta Mol Basis Dis. 2012;1822(5):639-49. doi: 10.1016/j.bbadis.2011.10.011.
- Furukawa H, Singh SK, Mancusso R, Gouaux E. Subunit arrangement and function in NMDA receptors. Nature. 2005;438(7065):185-92. doi: 10.1038/nature04089, PMID 16281028.

18. Paradis E, Clément S, Julien P, Ven Murthy MR. Lipoprotein lipase affects the survival and differentiation of neural cells exposed to very low-density lipoprotein. *J Biol Chem.* 2003;278(11):9698-705. doi: 10.1074/jbc.M208452200, PMID 12501246.
19. Azmi AS. Adopting network pharmacology for cancer drug discovery. *Curr Drug Discov Technol.* 2013;10(2):95-105. doi: 10.2174/1570163811310020002, PMID 23237672.
20. Jack Jr CR, Lowe VJ, Weigand SD, Wiste HJ, Senjem ML, Knopman DS, et al. Serial PIB and MRI in normal, mild cognitive impairment and Alzheimer's disease: implications for sequence of pathological events in Alzheimer's disease. *Brain.* 2009;132(5):1355-65. doi: 10.1093/brain/awp062, PMID 19339253.
21. Singh N, Pandey BR, Verma P. An overview of phytotherapeutic approach in prevention and treatment of Alzheimer's syndrome and dementia. *Int J Pharm Sci Drug Res.* 2011;3(3):162-72.
22. Alzheimer's Association. Alzheimer's disease facts and figures. *Alzheimers Dem.* 2014;10(2):e47-92.
23. Rafii MS, Aisen PS. Advances in Alzheimer's disease drug development. *BMC Med.* 2015;13(1):1-7.
24. Butler MS. The role of natural product chemistry in drug discovery. *J Nat Prod.* 2004;67(12):2141-53. doi: 10.1021/np040106y, PMID 15620274.
25. Khan I, Samad A, Khan AZ, Habtemariam S, Badshah A, Abdullah SM, et al. Molecular interactions of 4-acetoxy-plakinamine B with peripheral anionic and other catalytic subsites of the aromatic gorge of acetylcholinesterase: computational and structural insights. *Pharm Biol.* 2013;51(6):722-7. doi: 10.3109/13880209.2013.764329, PMID 23570516.
26. Richard T, Papastamoulis Y, Waffo-Teguo P, Monti JP. 3D NMR structure of a complex between the amyloid beta peptide (1-40) and the polyphenol ϵ -viniferin glucoside: implications in Alzheimer's disease. *Biochim Biophys Acta.* 2013;1830(11):5068-74. doi: 10.1016/j.bbagen.2013.06.031, PMID 23830862.
27. Crichton GE, Bryan J, Murphy KJ. Dietary antioxidants, cognitive function and dementia-a systematic review. *Plant Foods Hum Nutr.* 2013;68(3):279-92. doi: 10.1007/s11130-013-0370-0, PMID 23881465.
28. Devore EE, Grodstein F, van Rooij FJ, Hofman A, Stampfer MJ, Witteman JC, et al. Dietary antioxidants and long-term risk of dementia. *Arch Neurol.* 2010;67(7):819-25. doi: 10.1001/archneurol.2010.144, PMID 20625087.
29. Rodacka A, Strumillo J, Serafin E, Puchala M. Effect of resveratrol and tiron on the inactivation of glyceraldehyde-3-phosphate dehydrogenase induced by superoxide anion radical. *Curr Med Chem.* 2014;21(8):1061-9. doi: 10.2174/09298673113206660274, PMID 24059224.
30. Lakey-Beitia J, Berrocal R, Rao KS, Durant AA. Polyphenols as therapeutic molecules in Alzheimer's disease through modulating amyloid pathways. *Mol Neurobiol.* 2015;51(2):466-79. doi: 10.1007/s12035-014-8722-9, PMID 24826916.
31. Ramesh BN, Indi SS, Rao KS. Anti-amyloidogenic property of leaf aqueous extract of *Caesalpinia crista*. *Neurosci Lett.* 2010;475(2):110-4. doi: 10.1016/j.neulet.2010.03.062, PMID 20356566.
32. Berrocal R, Vasudevaraju P, Indi SS, Sambasiva Rao KR, Rao KS. *In vitro* evidence that an aqueous extract of *Centella asiatica* modulates α -synuclein aggregation dynamics. *J Alzheimers Dis.* 2014;39(2):457-65. doi: 10.3233/JAD-131187, PMID 24284367.
33. Pérez-Jiménez J, Neveu V, Vos F, Scalbert A. Systematic analysis of the content of 502 polyphenols in 452 foods and beverages: an application of the phenol-explorer database. *J Agric Food Chem.* 2010;58(8):4959-69. doi: 10.1021/jf100128b, PMID 20302342.
34. Mora-Pale M, Sanchez-Rodriguez SP, Linhardt RJ, Dordick JS, Koffas MA. Metabolic engineering and *in vitro* biosynthesis of phytochemicals and non-natural analogues. *Plant Sci.* 2013;210:10-24. doi: 10.1016/j.plantsci.2013.05.005, PMID 23849109.
35. Wang Y, Xia Z, Xu JR, Wang YX, Hou LN, Qiu Y, et al. α -mangostin, a polyphenolic xanthone derivative from mangosteen, attenuates β -amyloid oligomers-induced neurotoxicity by inhibiting amyloid aggregation. *Neuropharmacology.* 2012;62(2):871-81. doi: 10.1016/j.neuropharm.2011.09.016, PMID 21958557.
36. Morelli CF, Biagiotti M, Pappalardo VM, Rabuffetti M, Speranza G. Chemistry of α -mangostin. Studies on the semisynthesis of minor xanthenes from *Garcinia mangostana*. *Nat Prod Res.* 2015;29(8):750-5. doi: 10.1080/14786419.2014.986729, PMID 25482370.
37. Ibrahim MY, Hashim NM, Mohan S, Abdulla MA, Abdelwahab SI, Arbab IA, et al. α -mangostin from *Cratogeomys arborescens*: an *in vitro* and *in vivo* toxicological evaluation. *Arab J Chem.* 2015;8(1):129-37. doi: 10.1016/j.arabj.2013.11.017.
38. Buravlev EV, Shevchenko OG, Kutchin AV. Synthesis and membrane-protective activity of novel derivatives of α -mangostin at the C-4 position. *Bioorg Med Chem Lett.* 2015;25(4):826-9. doi: 10.1016/j.bmcl.2014.12.075, PMID 25592715.
39. Chajjaroenkul W, Na-Bangchang K. The *in vitro* antimalarial interaction of 9-hydroxycalabaxanthone and α -mangostin with mefloquine/artesunate. *Acta Parasitol.* 2014;60(1):105-11. doi: 10.1515/ap-2015-0013, PMID 26204026.
40. Choi M, Kim YM, Lee S, Chin YW, Lee C. Mangosteen xanthenes suppress hepatitis C virus genome replication. *Virus Genes.* 2014;49(2):208-22. doi: 10.1007/s11262-014-1098-0, PMID 24986787.
41. Koh JJ, Lin S, Aung TT, Lim F, Zou H, Bai Y, et al. Amino acid modified xanthone derivatives: novel, highly promising membrane-active antimicrobials for multidrug-resistant Gram-positive bacterial infections. *J Med Chem.* 2015;58(2):739-52. doi: 10.1021/jm501285x, PMID 25474410.
42. Shrestha S, Natarajan S, Park JH, Lee DY, Cho JG, Kim GS, et al. Potential neuroprotective flavonoid-based inhibitors of CDK5/p25 from *Rhus parviflora*. *Bioorg Med Chem Lett.* 2013;23(18):5150-4. doi: 10.1016/j.bmcl.2013.07.020, PMID 23927974.
43. Yang Z, Song Z, Xue W, Sheng J, Shu Z, Shi Y, et al. Synthesis and structure-activity relationship of nucleiferin derivatives as potential acetylcholinesterase inhibitors. *Med Chem Res.* 2014;23(6):3178-86. doi: 10.1007/s00044-013-0905-9.
44. Yang ZD, Zhang X, Du J, Ma ZJ, Guo F, Li S, et al. An aporphine alkaloid from *Nelumbo nucifera* as an acetylcholinesterase inhibitor and the primary investigation for structure-activity correlations. *Nat Prod Res.* 2012;26(5):387-92. doi: 10.1080/14786419.2010.487188, PMID 21732870.
45. Chatonnet A, Lockridge O. Comparison of butyrylcholinesterase and acetylcholinesterase. *Biochem J.* 1989;260(3):625-34. doi: 10.1042/bj2600625, PMID 2669736.
46. Gao D, Zhan CG. Modeling effects of oxyanion hole on the ester hydrolysis catalyzed by human cholinesterases. *J Phys Chem B.* 2005;109(48):23070-6. doi: 10.1021/jp053736x, PMID 16854005.
47. Bishara D, Harwood D. Safe prescribing of physical health medication in patients with dementia. *Int J Geriatr Psychiatry.* 2014;29(12):1230-41. doi: 10.1002/gps.4163, PMID 25092795.
48. Singh DB, Gupta MK, Kesharwani RK, Sagar M, Dwivedi S, Misra K. Molecular drug targets and therapies for Alzheimer's disease. *Transl Neurosci.* 2014;5(3):203-17. doi: 10.2478/s13380-014-0222-x.
49. Wong KY, Mercader AG, Saavedra LM, Honarparvar B, Romanelli GP, Duchowicz PR. QSAR analysis on tacrine-related acetylcholinesterase inhibitors. *J Biomed Sci.* 2014;21(1):84. doi: 10.1186/s12929-014-0084-0, PMID 25239202.
50. Qu J, Zhou Q, Du Y, Zhang W, Bai M, Zhang Z, et al. Rutin protects against cognitive deficits and brain damage in rats with chronic cerebral hypoperfusion. *Br J Pharmacol.* 2014;171(15):3702-15. doi: 10.1111/bph.12725, PMID 24758388.
51. Grosso C, Valentão P, Ferreres F, Andrade PB. The use of flavonoids in central nervous system disorders. *Curr Med Chem.* 2013;20(37):4694-719. doi: 10.2174/09298673113209990155, PMID 23834189.
52. Henry MS, Passmore AP, Todd S, McGuinness B, Craig D, Johnston JA. The development of effective biomarkers for Alzheimer's disease: a review. *Int J Geriatr Psychiatry.* 2013;28(4):331-40. doi: 10.1002/gps.3829, PMID 22674539.
53. Holmes C. Review: systemic inflammation and Alzheimer's disease. *Neuropathol Appl Neurobiol.* 2013;39(1):51-68. doi: 10.1111/j.1365-2990.2012.01307.x, PMID 23046210.
54. Li SY, Wang XB, Xie SS, Jiang N, Wang KD, Yao HQ, et al. Multifunctional tacrine-flavonoid hybrids with cholinergic, β -amyloid-reducing, and metal chelating properties for the treatment of Alzheimer's disease. *Eur J Med Chem.* 2013;69:632-46. doi: 10.1016/j.ejmech.2013.09.024, PMID 24095756.
55. Hyde C, Peters J, Bond M, Rogers G, Hoyle M, Anderson R, et al. Evolution of the evidence on the effectiveness and cost-effectiveness of acetylcholinesterase inhibitors and memantine for Alzheimer's disease: systematic review and economic model. *Age Ageing.* 2013;42(1):14-20. doi: 10.1093/ageing/afs165, PMID 23179169.
56. Anand P, Singh B. A review on cholinesterase inhibitors for Alzheimer's disease. *Arch Pharm Res.* 2013;36(4):375-99. doi: 10.1007/s12272-013-0036-3, PMID 23435942.
57. Struble RG, Ala T, Patrylo PR, Brewer GJ, Yan XX. Is brain amyloid production a cause or a result of dementia of the Alzheimer's type? *J Alzheimers Dis.* 2010;22(2):393-9. doi: 10.3233/JAD-2010-100846, PMID 20847431.
58. Luo W, Su YB, Hong C, Tian RG, Su LP, Wang YQ, et al. Design, synthesis and evaluation of novel 4-dimethylamine flavonoid derivatives as potential multi-functional anti-Alzheimer agents. *Bioorg Med Chem.* 2013;21(23):7275-82. doi: 10.1016/j.bmc.2013.09.061, PMID 24148835.
59. Goyal M, Grover S, Dhanjal JK, Goyal S, Tyagi C, Grover A. Molecular modelling studies on flavonoid derivatives as dual site inhibitors of human acetyl cholinesterase using 3D-QSAR, pharmacophore and high throughput screening approaches. *Med Chem Res.* 2014;23(4):2122-32. doi: 10.1007/s00044-013-0810-2.
60. Chakraborty S, Basu S. Insight into the anti-amyloidogenic activity of polyphenols and its application in virtual screening of phytochemical database. *Med Chem Res.* 2014;23(12):5141-8. doi: 10.1007/s00044-014-1081-2.
61. Lemkul JA, Bevan DR. Morin inhibits the early stages of amyloid β -peptide aggregation by altering tertiary and quaternary interactions to produce "off-pathway" structures. *Biochemistry.* 2012;51(30):5990-6009. doi: 10.1021/bi300113x, PMID 22762350.
62. Lemkul JA, Bevan DR. Destabilizing Alzheimer's A β 42 protofibrils with morin: mechanistic insights from molecular dynamics simulations. *Biochemistry.* 2010;49(18):3935-46. doi: 10.1021/bi1000855, PMID 20369844.
63. Remya C, Dileep KV, Tintu I, Variyar EJ, Sadasivan C. Design of potent inhibitors of acetylcholinesterase using morin as the starting compound. *Front Life Sci.* 2012;6(3-4):107-17. doi: 10.1080/21553769.2013.815137.
64. Duan S, Guan X, Lin R, Liu X, Yan Y, Lin R, et al. Silibinin inhibits acetylcholinesterase activity and amyloid β peptide aggregation: a dual-target drug for the treatment of Alzheimer's disease. *Neurobiol Aging.* 2015;36(5):1792-807. doi: 10.1016/j.neurobiolaging.2015.02.002, PMID 25771396.
65. Boratyński PJ. Dimeric cinchona alkaloids. *Mol Divers.* 2015;19(2):385-422. doi: 10.1007/s11030-014-9563-1, PMID 25586655.
66. Larsson S, Rønsted N. Reviewing Colchicaceae alkaloids—perspectives of evolution on medicinal chemistry. *Curr Top Med Chem.* 2014;14(2):274-89. doi: 10.2174/1568026613666131216110417, PMID 24359194.

67. Vale VV, Vilhena TC, Trindade RC, Ferreira MR, Percário S, Soares LF, et al. Anti-malarial ether and toxicity assessment of *Himatanthus articulatus*, a plant used to treat malaria in the Brazilian Amazon. *Malar J*. 2015;14(1):132. doi: 10.1186/s12936-015-0643-1, PMID 25888719.
68. Kong DG, Zhao Y, Li GH, Chen BJ, Wang XN, Zhou HL, et al. The genus *Litsea* in traditional Chinese medicine: an ethnomedical, phytochemical and pharmacological review. *J Ethnopharmacol*. 2015;164:256-64. doi: 10.1016/j.jep.2015.02.020, PMID 25698244.
69. Jiang WW, Su J, Wu XD, He J, Peng LY, Cheng X, et al. Geissoschizine methyl ether N-oxide, a new alkaloid with antiacetyl cholinesterase activity from *Uncariahynchophylla*. *Nat Prod Res*. 2015;29(9):842-7. doi: 10.1080/14786419.2014.989847, PMID 25496282.
70. Liu W, Shi X, Yang Y, Cheng X, Liu Q, Han H, et al. *In vitro* and *in vivo* metabolism and inhibitory activities of vasicine, a potent acetylcholinesterase and butyrylcholinesterase inhibitor. *PLOS ONE*. 2015;10(4):e0122366. doi: 10.1371/journal.pone.0122366, PMID 25849329.
71. Liew SY, Khaw KY, Murugaiyah V, Looi CY, Wong YL, Mustafa MR, et al. Natural indole butyrylcholinesterase inhibitors from *Nauclea officinalis*. *Phytomedicine*. 2015;22(1):45-8. doi: 10.1016/j.phymed.2014.11.003, PMID 25636869.
72. Shaikh S, Zainab T, Shakil S, Rizvi SM. A neuroinformatics study to compare inhibition efficiency of three natural ligands (Fawcettimine, Cernuine and Lycodine) against human brain acetylcholinesterase. *Network*. 2015;26(1):25-34. doi: 10.3109/0954898X.2014.994145, PMID 25611730.
73. Konrath EL, Passos CD, Klein LC, Henriques AT. Alkaloids as a source of potential anticholinesterase inhibitors for the treatment of Alzheimer's disease. *J Pharm Pharmacol*. 2013;65(12):1701-25. doi: 10.1111/jphp.12090, PMID 24236981.
74. Huang HJ, Lee CC, Chen CY. Lead discovery for Alzheimer's disease related target protein RbAp48 from traditional Chinese medicine. *BioMed Res Int*. 2014;2014:764946. doi: 10.1155/2014/764946, PMID 25165715.
75. Makowska J, Szczesny D, Lichucka A, Gieldon A, Chmurzyński L, Kalisz R. Preliminary studies on trigonelline as potential anti-Alzheimer disease agent: determination by hydrophilic interaction liquid chromatography and modeling of interactions with beta-amyloid. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2014;968:101-4. doi: 10.1016/j.jchromb.2013.12.001, PMID 24374010.
76. Cortes N, Alvarez R, Osorio EH, Alzate F, Berkov S, Osorio E. Alkaloid metabolite profiles by GC/MS and acetylcholinesterase inhibitory activities with binding-mode predictions of five Amaryllidaceae plants. *J Pharm Biomed Anal*. 2015;102:222-8. doi: 10.1016/j.jpba.2014.09.022, PMID 25305596.
77. Cahliková L, Valterová I, Macáková K, Opletal L. Analysis of Amaryllidaceae alkaloids from *Zephyranthes grandiflora* by GC/MS and their cholinesterase activity. *Rev Bras Farmacognosia*. 2011;21(4):575-80. doi: 10.1590/S0102-695X2011005000089.
78. Kulhánková A, Cahliková L, Novák Z, Macáková K, Kuneš J, Opletal L. Alkaloids from *Zephyranthes robusta* Baker and their acetylcholinesterase- and butyrylcholinesterase-inhibitory activity. *Chem Biodivers*. 2013;10(6):1120-7. doi: 10.1002/cbdv.201200144, PMID 23776027.
79. Larsen MM, Adersen A, Davis AP, Lledó MD, Jäger AK, Rønsted N. Using a phylogenetic approach to selection of target plants in drug discovery of acetylcholinesterase inhibiting alkaloids in Amaryllidaceae tribe Galantheae. *Biochem Syst Ecol*. 2010;38(5):1026-34. doi: 10.1016/j.bse.2010.10.005.
80. Jahn S, Seiwert B, Kretzing S, Abraham G, Regenthal R, Karst U. Metabolic studies of the Amaryllidaceae alkaloids galantamine and lycorine based on electrochemical simulation in addition to *in vivo* and *in vitro* models. *Anal Chim Acta*. 2012;756:60-72. doi: 10.1016/j.aca.2012.10.042, PMID 23176740.
81. Sharma V, Hussain S, Bakshi M, Bhat N, Saxena AK. *In vitro* cytotoxic activity of leaves extracts of *Holarrhena antidysenterica* against some human cancer cell lines. *Indian J Biochem Biophys*. 2014;51(1):46-51. PMID 24791416.
82. Chusri S, Na-Phatthalung P, Siriyong T, Paosen S, Voravuthikunchai SP. *Holarrhena antidysenterica* as a resistance modifying agent against *Acinetobacter baumannii*: its effects on bacterial outer membrane permeability and efflux pumps. *Microbiol Res*. 2014;169(5-6):417-24. doi: 10.1016/j.micres.2013.09.004, PMID 24103863.
83. Dua VK, Verma G, Singh B, Rajan A, Bagai U, Agarwal DD, et al. Anti-malarial property of steroidal alkaloid conessine isolated from the bark of *Holarrhena antidysenterica*. *Malar J*. 2013;12(1):194. doi: 10.1186/1475-2875-12-194, PMID 23758861.
84. Kadir MF, Bin Sayeed MS, Mia MM. Ethnopharmacological survey of medicinal plants used by traditional healers in Bangladesh for gastrointestinal disorders. *J Ethnopharmacol*. 2013;147(1):148-56. doi: 10.1016/j.jep.2013.02.023, PMID 23458917.
85. Yang ZD, Duan DZ, Xue WW, Yao XJ, Li S. Steroidal alkaloids from *Holarrhena antidysenterica* as acetylcholinesterase inhibitors and the investigation for structure-activity relationships. *Life Sci*. 2012;90(23-24):929-33. doi: 10.1016/j.lfs.2012.04.017, PMID 22569298.
86. Ul-Haq Z, Mahmood U, Jehangir B. Ligand-based 3D-QSAR studies of physostigmine analogues as acetylcholinesterase inhibitors. *Chem Biol Drug Des*. 2009;74(6):571-81. doi: 10.1111/j.1747-0285.2009.00897.x, PMID 19843075.
87. Plaschke K, Müller AK, Kopitz J. Surgery-induced changes in rat IL-1 β and acetylcholine metabolism: role of physostigmine. *Clin Exp Pharmacol Physiol*. 2014;41(9):663-70. doi: 10.1111/1440-1681.12267, PMID 24890001.
88. Killi UK, Wsol V, Soukup O, Kuca K, Winder M, Tobin G. *In vitro* functional interactions of acetylcholine esterase inhibitors and muscarinic receptor antagonists in the urinary bladder of the rat. *Clin Exp Pharmacol Physiol*. 2014;41(2):139-46. doi: 10.1111/1440-1681.12191, PMID 24341923.
89. Yokota SI, Nakamura K, Ando M, Kamei H, Hakuno F, Takahashi SI, et al. Acetylcholinesterase (AChE) inhibition aggravates fasting-induced triglyceride accumulation in the mouse liver. *FEBS Open Bio*. 2014;4:905-14. doi: 10.1016/j.fob.2014.10.009, PMID 25383314.
90. Shaikh S, Verma A, Siddiqui S, Ahmad S S, MD Rizvi S, Shakil S, Biswas D, Singh D, H Siddiqui M, Shakil S, Tabrez S. Current acetylcholinesterase-inhibitors: a neuroinformatics perspective. *CNS and Neurological Disorders-Drug Targets (Formerly Current Drug Targets-CNS and Neurological Disorders)*. 2014;13(3):391-401.
91. Amat-Ur-Rasool H, Ahmed M. Designing second generation anti-alzheimer compounds as inhibitors of human acetylcholinesterase: computational screening of synthetic molecules and dietary phytochemicals. *PLOS ONE*. 2015;10(9):e0136509. doi: 10.1371/journal.pone.0136509, PMID 26325402.
92. Scotti L, Scotti MT, Ishiki H, Junior FJ, dos Santos PF, Tavares JF, et al. Prediction of anticancer activity of diterpenes isolated from the Paraíba flora through a PLS model and molecular surfaces. *Nat Prod Commun*. 2014;9(5):1934578X1400900503:609-12. doi: 10.1177/1934578X1400900503, PMID 25026699.
93. Scotti L, Tavares JF, Silva MS, Falcão EV, Silva GC, Soares MT, et al. Chemotaxonomy of three genera of the Annonaceae family using self-organizing maps and ¹³C NMR data of diterpenes. *Quim Nova*. 2012;35(11):2146-52. doi: 10.1590/S0100-40422012001100008.
94. Scotti MT, Fernandes MB, Ferreira MJ, Emerenciano VP. Quantitative structure-activity relationship of sesquiterpene lactones with cytotoxic activity. *Bioorg Med Chem*. 2007;15(8):2927-34. doi: 10.1016/j.bmc.2007.02.005, PMID 17336532.
95. Scotti MT, Emerenciano V, Ferreira MJ, Scotti L, Stefani R, Da Silva MS, et al. Self-organizing maps of molecular descriptors for sesquiterpene lactones and their application to the chemotaxonomy of the Asteraceae family. *Molecules*. 2012;17(4):4684-702. doi: 10.3390/molecules17044684, PMID 22522398.
96. Galipoğlu M, Erdal MS, Güngör S. Biopolymer-based transdermal films of donepezil as an alternative delivery approach in Alzheimer's disease treatment. *AAPS PharmSciTech*. 2015;16(2):284-92. doi: 10.1208/s12249-014-0224-6, PMID 25273029.
97. Guo Q, Ma X, Wei S, Qiu D, Wilson IW, Wu P, et al. De novo transcriptome sequencing and digital gene expression analysis predict biosynthetic pathway of rhynchophylline and isorhynchophylline from *Uncariahynchophylla*, a non-model plant with potent anti-alzheimer's properties. *BMC Genomics*. 2014;15(1):1-6.
98. Xie H, Wang JR, Yau LF, Liu Y, Liu L, Han QB, et al. Quantitative analysis of the flavonoid glycosides and terpene trilactones in the extract of *Ginkgo biloba* and evaluation of their inhibitory activity towards fibril formation of β -amyloid peptide. *Molecules*. 2014;19(4):4466-78. doi: 10.3390/molecules19044466, PMID 24727418.
99. Yin Y, Zhang Y, Li P, Ma H, Zhang X, Pan G, et al. 2014. Pharmaceutical composition useful for treating Alzheimer's syndrome, comprises *Lycium barbarum* polysaccharide-III, medlar polysaccharideIV, resveratrol, terpene-3-beta-alcohol, Astragalus polysaccharide A2 and motherwort saponin A. CN103751207-A, CN103751207-A 30 Apr 2014 A61K-031/715 201448.
100. Bidon-Chanal A, Fuertes A, Alonso D, Pérez DI, Martínez A, Luque FJ, et al. Evidence for a new binding mode to GSK-3: allosteric regulation by the marine compound palinurin. *Eur J Med Chem*. 2013;60:479-89. doi: 10.1016/j.ejmech.2012.12.014, PMID 23354070.
101. Sivaprakasam P, Han X, Civiello RL, Jacutin-Porte S, Kish K, Pokross M, et al. Discovery of new acylaminopyridines as GSK-3 inhibitors by a structure guided in-depth exploration of chemical space around a pyrrolopyridinone core. *Bioorg Med Chem Lett*. 2015;25(9):1856-63. doi: 10.1016/j.bmcl.2015.03.046, PMID 25845281.
102. Ye Q, Mao W, Zhou Y, Xu L, Li Q, Gao Y, et al. Synthesis and biological evaluation of 3-([1, 2, 4] triazololo [4, 3-a] pyridin-3-yl)-4-(indol-3-yl)-maleimides as potent, selective GSK-3 β inhibitors and neuroprotective agents. *Bioorg Med Chem*. 2015;23(5):1179-88. doi: 10.1016/j.bmc.2014.12.026, PMID 25662701.
103. Kalakech H, Hibert P, Prunier-Mirebeau D, Tamareille S, Letournel F, Macchi L, et al. RISK and SAFE signaling pathway involvement in apolipoprotein AI-induced cardioprotection. *PLOS ONE*. 2014;9(9):e107950. doi: 10.1371/journal.pone.0107950, PMID 25237809.
104. Feng H, Yu Z, Tian Y, Lee YY, Li MS, Go MY, et al. A CCRK-EZH2 epigenetic circuitry drives hepatocarcinogenesis and associates with tumor recurrence and poor survival of patients. *J Hepatol*. 2015;62(5):1100-11. doi: 10.1016/j.jhep.2014.11.040, PMID 25500144.

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